

## Review



**Cite this article:** Aguirre J, Catalán P, Cuesta JA, Manrubia S. 2018 On the networked architecture of genotype spaces and its critical effects on molecular evolution. *Open Biol.* **8**: 180069.

<http://dx.doi.org/10.1098/rsob.180069>

Received: 18 April 2018

Accepted: 12 June 2018

**Subject Area:**

systems biology

**Keywords:**

genotype–phenotype map, network-of-networks, adaptive multiscapes, molecular promiscuity, phenotypic plasticity, punctuated dynamics

**Author for correspondence:**

Susanna Manrubia

e-mail: [smanrubia@cnb.csic.es](mailto:smanrubia@cnb.csic.es)

# On the networked architecture of genotype spaces and its critical effects on molecular evolution

Jacobo Aguirre<sup>1,2</sup>, Pablo Catalán<sup>1,3</sup>, José A. Cuesta<sup>1,3,4,5</sup>  
and Susanna Manrubia<sup>1,2</sup>

<sup>1</sup>Grupo Interdisciplinar de Sistemas Complejos (GISC), Madrid, Spain

<sup>2</sup>Programa de Biología de Sistemas, Centro Nacional de Biotecnología (CSIC), Madrid, Spain

<sup>3</sup>Departamento de Matemáticas, Universidad Carlos III de Madrid, Leganés, Madrid, Spain

<sup>4</sup>Instituto de Biocomputación y Física de Sistemas Complejos (BIFI), Universidad de Zaragoza, Zaragoza, Spain

<sup>5</sup>UC3M-BS Institute of Financial Big Data (IFiBiD), Universidad Carlos III de Madrid, Getafe, Madrid, Spain

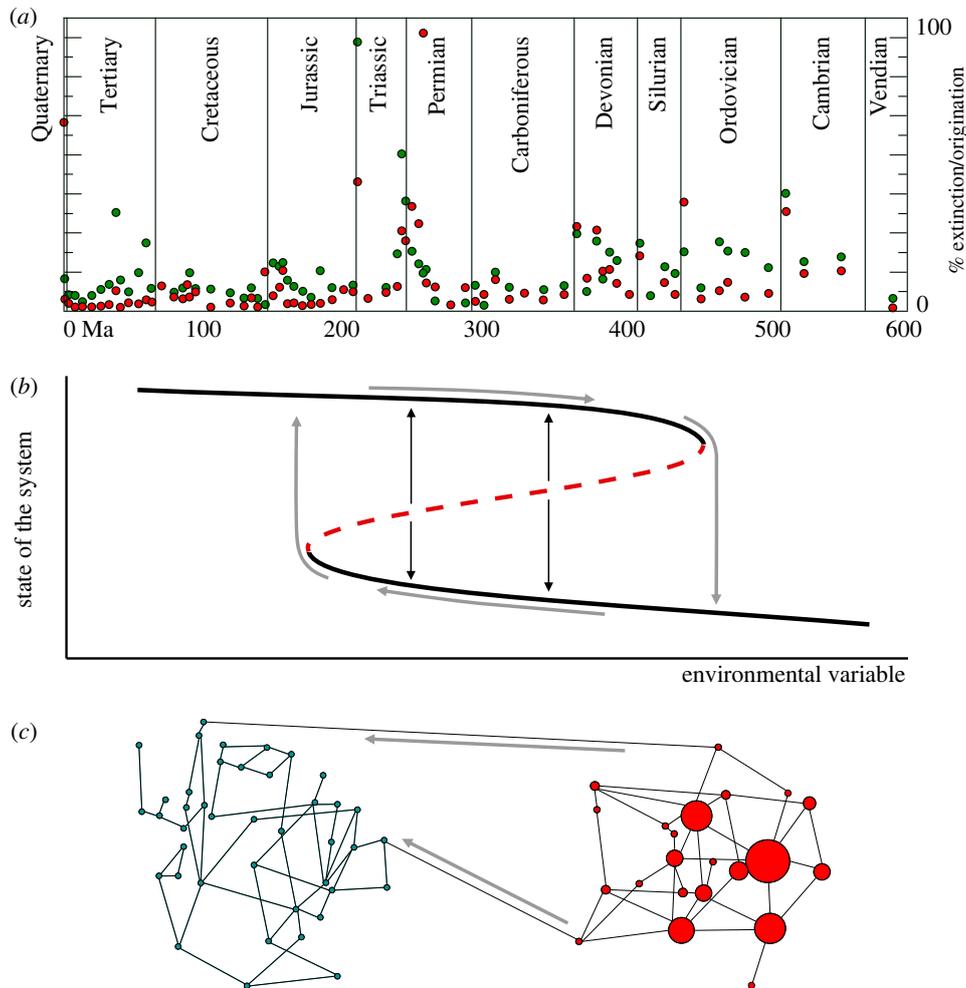
JA, 0000-0003-2196-5103; PC, 0000-0003-2826-4684; SM, 0000-0003-0134-2785

Evolutionary dynamics is often viewed as a subtle process of change accumulation that causes a divergence among organisms and their genomes. However, this interpretation is an inheritance of a gradualistic view that has been challenged at the macroevolutionary, ecological and molecular level. Actually, when the complex architecture of genotype spaces is taken into account, the evolutionary dynamics of molecular populations becomes intrinsically non-uniform, sharing deep qualitative and quantitative similarities with slowly driven physical systems: nonlinear responses analogous to critical transitions, sudden state changes or hysteresis, among others. Furthermore, the phenotypic plasticity inherent to genotypes transforms classical fitness landscapes into multiscapes where adaptation in response to an environmental change may be very fast. The quantitative nature of adaptive molecular processes is deeply dependent on a network-of-networks multi-layered structure of the map from genotype to function that we begin to unveil.

## 1. Introduction

Gradualism posits that any profound change in nature is the result of minor cumulative modifications due to the action of slow but sustained processes. First proposed in the framework of Geology at the end of the eighteenth century by James Hutton, gradualism underlies Charles Lyell's theory of uniformitarianism [1], which formed one of the conceptual pillars of Charles Darwin's evolutionary theory soon after [2]. Ever since, gradualism has been a powerful concept in the qualitative interpretation of evolutionary change.

The gradualistic view of evolution has been challenged at the macro- (fossil record), meso- (ecological) and micro- (molecular) scales. In the 1970s, analyses of data in the fossil record revealed an unanticipated pattern of evolutionary stasis in the morphological change of species that was punctuated by sudden jumps, leading to the theory of punctuated equilibria [3]. The mechanistic models proposed to generate that dynamical pattern are not unique, though the endogenous organization of the biosphere may have played a main role [4,5]. At present, punctuated equilibrium is understood as an alternation of periods with insignificant change (stasis) punctuated by rapid speciation, which may, however, extend over a few hundred thousand years and result from complex evolutionary dynamics [6]. Analogies between macroevolution and evolutionary ecology were suggested on the basis that the degree of complexity observed in the spatial and temporal organization of both systems might be reflecting a network-like organization close to critical points [7], the latter resulting from a combination of external drivers and internal adaptive responses.



**Figure 1.** Punctuated behaviour in macroevolution, ecology and molecular dynamics. (a) Non-uniform pattern of extinctions (red symbols) and originations (green symbols) in the last 610 Myr (0 is present). Each point corresponds to a geological epoch, vertical lines separate geological periods, as indicated. The vertical axis gives the percentage of extinction/origination per estimated diversity at each epoch and per million years. Data from [12], geological epochs and periods as in [13]. (b) Minor changes in environmental variables might cause large, nonlinear responses in the state of a variety of systems. In some cases, two stable solutions (black curves) coexist with an unstable solution (red curve) for a range of values of a control parameter. The trajectories of systems might follow the path indicated by the grey arrows as that parameter increases, suffering a sudden jump from the upper to the lower branch. Hysteretic behaviour appears and prevents the recovery of the initial state when the environmental variable is reverted. When the system is initiated close to the unstable branch, it may attain any of the two possible stable solutions (black thin arrows). (c) In the genotype space, nodes represent genotypes and links correspond to single mutational moves. Heterogeneous molecular populations contain a set of genotypes with variable abundances, the latter represented through circle size. Fitter regions in genotype space might be difficult to find if there are few mutational incoming pathways (grey arrows). The population might be trapped in the red phenotype for a relatively long time (stasis) when compared with the transition to the new state once suitable mutations have appeared (punctuation).

Research in this century has unveiled a large number of cases where smooth environmental changes may indeed trigger sudden and irreversible ecological responses [8,9]. The complex interaction between natural systems and varying environments remains an open question of critical relevance. The factors that make ecosystems respond smoothly or drastically to a weakly evolving environment have attracted special interest, as there are direct implications in the relationship between humans and a changing biosphere that could eventually reach a hazardous tipping point [8,10,11].

The formal description of non-uniform dynamics in natural systems is advancing concomitantly with the number of examples supporting and clarifying the theoretical framework (figure 1). Shifts in ecosystems have been formally described as bifurcations leading to hysteretic behaviour and also as critical transitions. Analogous to fluctuations close to critical points, the so-called early warning signals can anticipate such catastrophic responses [14]. Empirical evidence of this phenomenon with a single species has been described in laboratory

populations of yeast [15], while there is a variety of well-documented examples in ecology, such as the hysteretic loss and recovery of charophyte vegetation at lake Veluwe [16], the desertification of the Sahara [9,17], the loss of transparency in shallow lakes [18] or the dynamics of woodlands in Tanzania [19]. A thorough description of this phenomenology is a hard task, as it involves a wide variety of time scales and biological levels—many of them organized as complex networks—that interact in a complex manner [20]. At the molecular level, the architecture of the genotype–phenotype map entails non-uniform evolutionary dynamics [21]. In particular, it has been shown that the steady accumulation of point mutations under a selective pressure acting on the phenotype yields population dynamics characterized by stasis (when sequences explore neutral regions) punctuated by phenotypic changes (when a fitter phenotype is found) [22]. Smooth changes at the level of sequences do not preclude sudden adaptive changes at the level of function: well-motivated models support that, like the state of ecosystems, changes in genomic

composition might be sudden, irreversible and unavoidable [23]. These dynamics have been also documented in the *in vivo* evolution of a virus, influenza A, which shows a seasonal pattern where expansion of genotypic diversity predates the finding and fixation of strains with novel antigenic properties that escape immune detection [24,25].

Despite mounting evidence, the long tradition of relating small changes in sequences to gradual changes in organisms and populations persists, often in a tacit way. A significant example is Wright's adaptive landscape [26], which appears as a direct consequence of gradualistic thought and counts among the most powerful metaphors in Biology, one that has conditioned evolutionary thinking for almost a century [27]. Indeed, the image of a relatively smooth landscape, where populations adapt by going uphill, are trapped in mountain peaks and remain isolated from other possibly higher fitness maxima by deep valleys, often appears as the way in which adaptation proceeds. This picture implies a smooth and continuous genotype-to-phenotype (GP) map and a space of low dimensionality. Thanks to advances in our knowledge of the molecular structure of populations, we now know of important elements missing in most theoretical adaptive landscapes. For example, genotypes of similar fitness are found to form extensive networks that occasionally traverse the genotype space, especially in spaces of high dimensionality [28]. The GP map actually entails a many-to-many correspondence: genotypes are plastic and may yield different phenotypes when expressed in different environments. This latter case seems to be much more common than previously thought, meaning that the co-option of promiscuous, secondary gene functions [29] is likely a common adaptive mechanism. From a formal viewpoint, therefore, the complexity of the GP map implies that fitness landscapes should be visualized as high-dimensional and interwoven sets of networks that unfold into multiple layers under environmental change [30]. New techniques, in particular the use of deep sequencing and powerful massive ways to evaluate the fitness of individual genotypes, represent a breakthrough in the empirical characterization of the complex genotype-to-phenotype-to-function relationship [31,32]. Interestingly, the network-of-networks structure of genotype spaces described in realistic, though artificial, models is also emerging in empirical characterizations of the diversity of molecular populations [33].

Adaptive evolutionary systems, such as large-scale evolution, ecology or (molecular) populations, share deep analogies that can be likely ascribed to their networked architecture plus a non-trivial relationship between exogenous drivers and endogenous responses. In this review, we will focus on molecular dynamics, which is the least studied of those three profoundly entangled levels of description of the evolutionary process. The architecture of genotype spaces and the dynamics of evolving molecular populations are two sides of the same coin. The heterogeneous structure of genotype spaces and its apparently hierarchical organization as a multilayered network of networks explains, among others, punctuated dynamics [22], drift and switch transitions [24], genomic shifts [23] or Waddington's genetic assimilation [30,34].

## 2. Genotype networks

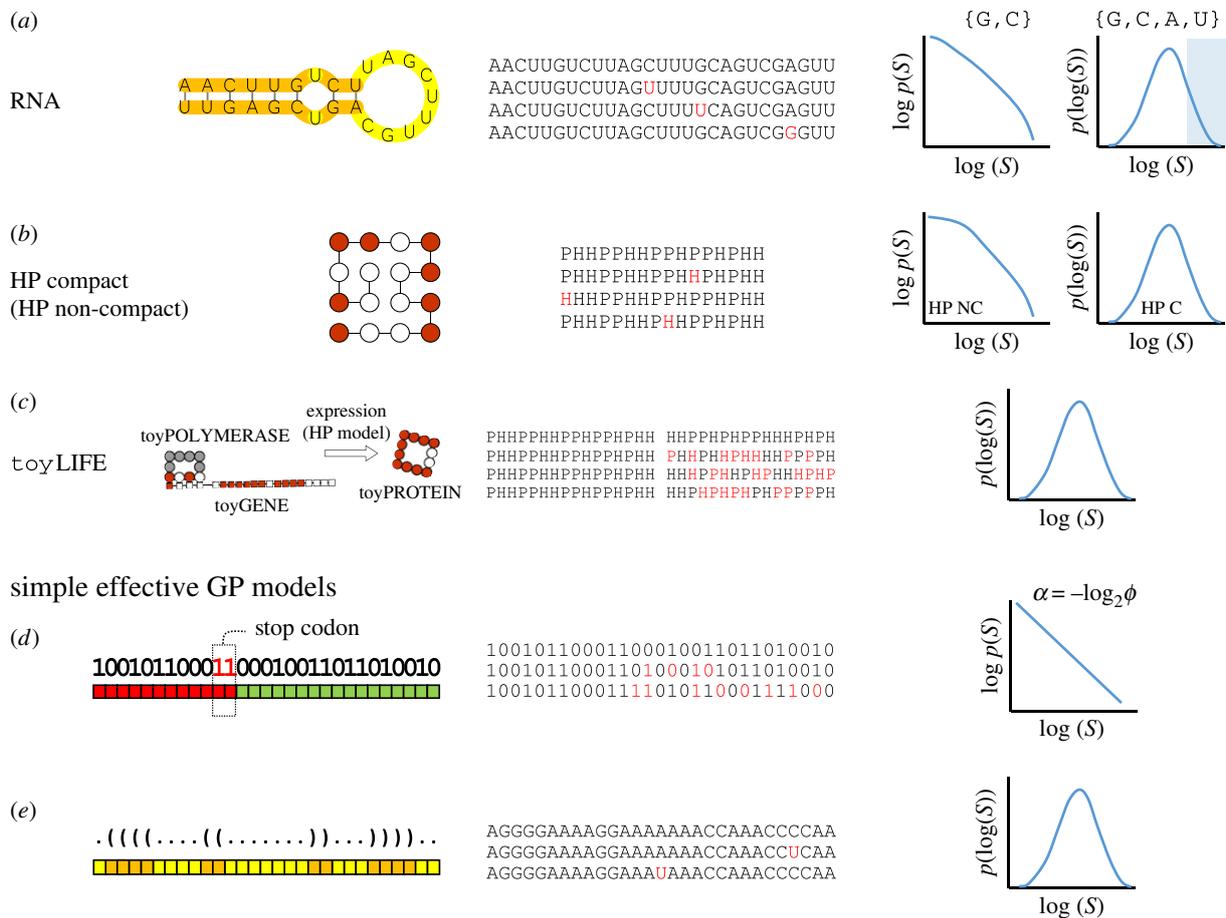
Kimura [35,36] introduced the concept of neutral evolution in order to explain why many mutations observed in RNA,

DNA or proteins do not affect fitness. Neutrality implies that the GP map is not one-to-one, but many-to-one, consistently explaining the high level of polymorphism observed in natural populations. Soon after Kimura's seminal work, navigability was hypothesized as an essential requirement to guarantee the evolvability of molecular populations [37]. Usually, navigability is believed to rely on the existence of sufficiently large *neutral networks* (NNs) of genotypes [38] since these should permit the neutral drift of populations and a sustained exploration of alternative phenotypes without a detrimental decrease in fitness. An NN is formed by all genotypes that map into the same phenotype. As fitness is linked to phenotype, all genotypes in an NN are implicitly assumed to have the same fitness. Genotypes are the nodes of such networks, and links correspond to single mutational moves. In its simplest and most popular definition, a mutational move stands for a point mutation. Neutral networks can have one or several connected components. Navigability on NNs has been subsequently identified as a robust property of computational models [22,39–41] and natural molecular populations [25,42–44].

The actual set of genotypes visited by an evolving population, however, is rarely neutral. Nearly neutral mutations are common in finite populations [45], augmenting their adaptive ability. In fact, any finite mutation rate entails that populations are heterogeneous in sequence, phenotype and function, such that the potential set of genotypes of a population includes genotypes of different fitness, which constitute the actual navigable network. In certain cases, as for ensembles of fast mutating replicators such as quasi-species [46,47], the maintenance of a large phenotypic diversity and the permanent exploration of the genome space become critical survival strategies [48]. We will call *genotype network* the network of visited genotypes and, by extension, any potentially navigable network in the space of genomes, regardless of the fitness or phenotype of its nodes.

### 2.1. Neutral networks in computational genotype–phenotype maps

Neutral networks have been quantitatively characterized in a number of computational GP maps (figure 2). RNA sequences fold into a minimum free energy secondary structure that we can take as a proxy for its phenotype [38,56]. Given a sequence length, the number of minimum free energy secondary structures is much smaller than the number of sequences, leading to large NNs [38,50,57–61]. In models of protein structure, such as the HP model [62], proteins are formed by strings of two amino acids: hydrophobic (H) and polar (P). As in RNA, this sequence will fold into a minimum free energy structure, and there are many more sequences than structures [51,63–65]. In a completely different model, gene regulatory networks possess an evolvable architecture [66] that gives rise to several temporal gene expression patterns, which represent the phenotype. Again, many interaction topologies representing the genotype give rise to a much smaller number of gene expression patterns [28,67]. Neutral networks also appear in metabolic processes. If we consider the genotype as a list of enzymatic reactions and the phenotype as the set of metabolic sources on which an organism can survive, it is found that many genotypes can actually survive in a set of environments [41,68–70]. Finally, NNs have also been observed in



**Figure 2.** Some examples of simple GP maps. For each model, and from left to right, we depict an example phenotype, some of the sequences in its neutral network (mutations that do not change the phenotype are highlighted in red), and the schematic functional form of the probability distribution  $p(S)$  of phenotypes sizes  $S$  found in computational or analytical studies. (a) RNA sequence-to-minimum-free-energy secondary structure. Mutations that do not disrupt the secondary structure appear with different probability in loops or stacks. In two-letter alphabets, the distribution of phenotype sizes is compatible with a power-law function [49], while in four-letter alphabets  $p(S)$  is well fit by a lognormal distribution [50]. For long sequences, only the right-most part of  $p(S)$  can be seen under random sampling of the genotype space [50] (shaded). (b) The HP model, in its compact (as in the figure) or non-compact versions, has been studied as a model for protein folding. In non-compact versions, the distribution  $p(S)$  has a maximum at  $S = 1$  and decays with a fat tail [51], while in compact versions  $p(S)$  resembles a lognormal distribution [52]. (c)  $\text{toyLIFE}$  is a minimal model with several levels. HP-like sequences are read and translated to proteins that interact through analogous rules to break metabolites. The  $p(S)$  of  $\text{toyLIFE}$  is compatible with a lognormal distribution [53]. (d,e) Effective models where phenotype is defined in relation to the composition of sequences allow to analytically calculate the functional form of  $p(S)$ . Two examples are (d) Fibonacci's model [54], where  $p(S)$  follows a power-law distribution and (e) an RNA-inspired model [55] which yields a lognormal distribution of  $p(S)$ .

complex models that include cellular population dynamics and several levels from genotype to phenotype [71], in more abstract GP maps, such as the polyomino model of polymer self-assembly [72,73],  $\text{toyLIFE}$ —a multilevel model of a simplified cellular biology [53,74]—and in simplified combinatorial models [54,55].

Most NNs studied in the literature share a remarkable number of structural properties [28,75]:

1. Most phenotypes are rare, and only a few of them are very common. Specifically, the probability of finding a phenotype when sampling uniformly at random among all of them follows a lognormal distribution for a wide variety of models [50,55,74] and a power law for some special cases [49,54,55]. Therefore, a small fraction of the largest phenotypes contains most genotypes, such that in practice those are the only ones visible to natural selection [50,59,76]; together with the asymmetry in the mutual accessibility of two phenotypes [58,77], that property causes a form of (entropic) trapping in genotype space [74,76,78,79].

2. The degree of a node in an NN, defined as the number of one-mutant neighbours that belong to the same NN (aka its genotypic robustness), is a heterogeneous quantity, although its distribution is often unimodal [28,39,61]. Additionally, the average degree of an NN is proportional to the logarithm of the size of the network [55,60,61,80].
3. These NNs are assortative, at least for phenotypes defined through minimum-energy principles [61,81,82]. In an assortative network, genotypes are connected to other genotypes of similar degree, and this correlation in genotypic robustness causes canalization [83], leads to phenotypic entrapment [79] and enhances evolvability [80].
4. Neutral networks of common phenotypes percolate genotype space. In other words, we can find two genotypes expressing the same phenotype with a sequence similarity comparable to that of two randomly chosen genotypes [28,84].
5. Most large phenotypes are one mutation away from each other, such that genotypes yielding every common phenotype can be found at the boundary of any large NN

[28,53,65,84]. As a result, the search for new phenotypes among common ones is a fast process.

The space of genotypes can be depicted in this context by a number of interconnecting NNs when each node is projected in a horizontal (quasi-) neutral layer whose vertical position represents its fitness value. In this multilayer perspective [85,86], intralayer connections between individual nodes represent neutral mutations, while interlayer connections represent mutations that beneficially (upwards) or deleteriously (downwards) affect fitness [87]. It is, however, important to keep in mind that this representation is suitable only if the GP map is approximated as a many-to-one relationship, since it fails to include the frequent correspondence between one genotype and several possible (environment dependent) phenotypes, as will be discussed in §§5 and 6.

## 2.2. Genotype networks in genotype-to-function maps

The GP map is at best a toy representation of the relationship between genotype and function, though it hopefully captures some of its statistical properties. Computational studies suggest that structural properties of GP maps are largely independent of the precise definition of phenotype [88,89] and of details of specific models [28,75], and data to assess whether GP maps are a sufficiently accurate representation of genotype-to-function maps—which represent a qualitative step forward—is mounting. Advances in experimental techniques have allowed to study the structure of the genotype-to-fitness mapping through either experimental evolution studies [33,90–93] or high-throughput data [32,44,94]. The resulting experimental fitness landscapes confirm and extend the picture of molecular evolution gained through the computational study of simple GP maps, showing the presence of many quasi-neutral (eventually navigable) regions [95] and decaying correlations between phenotypes as the mutational distance increases [96]. Natural fitness landscapes have an intermediate degree of ruggedness, they are neither smooth nor random, therefore revealing an important role of epistasis in shaping the topological properties of genotype networks and in defining eventually accessible genomic pathways for molecular adaptation [44,90,97,98].

Fitness landscapes have been theoretically explored through models where phenotypes need not be explicitly defined and, instead, a fitness value is associated with each genotype. This representation is closer to data retrieved through empirical evolutionary experiments. The NK model [99] has proved to be especially useful to generate an underlying landscape with realistic degrees of ruggedness [24,92]. Furthermore, it is relatively simple, only depending on two parameters—the length of the sequence  $N$  and the level of ruggedness  $K$ —but versatile enough to model fitness landscapes with natural properties such as epistasis, multiple fitness peaks and local optima [100].

It turns out that topological differences between genotype networks, obtained through data that map genotype to function, and NNs, as described in the previous subsection, are only cosmetic. It can be shown that spaces of genotypes endowed with the structure of the NK model are also organized as a network of networks, that is, as a set of genotype networks qualitatively equivalent to NNs connected through a limited number of pathways [101]. The structural properties of genotype networks, visualized as a multilayered network

of networks, define a particular class of dynamics for populations evolving on such architecture.

The following sections are devoted to the not yet fully understood interaction between the topology of genotype networks and the evolutionary dynamics of heterogeneous populations—at least from the formal viewpoint of dynamical systems. We begin by synthesizing current evidence to demonstrate that three different dynamical situations (competitive transitions between different regions of an NN [102], punctuated molecular adaptation [22] and genomic shifts under varying environments [23]) can be described within a unique conceptual and theoretical framework. In subsequent sections, we will show how the latter framework can be extended to include the many-to-many inherent structure of GP maps and environmental changes.

## 3. Population dynamics on neutral networks

In order to describe mathematically the evolution of heterogeneous populations on NNs, let us recall that many dynamical processes occurring on a network can be expressed as

$$\mathbf{n}(t) = \mathbf{M}\mathbf{n}(t-1) = \mathbf{M}^t\mathbf{n}(0), \quad (3.1)$$

where  $\mathbf{n}(t)$  is a vector whose components are the population of individuals at each node at time  $t$  and  $\mathbf{M}$  is an evolution matrix that contains the particulars of the dynamical process (see box 1).

For the sake of illustration, let us start by considering a simple fitness landscape with a single viable phenotype. The genotypes yielding the latter constitute an NN and all remaining genotypes have zero fitness. Consider genotypes as sequences of length  $l$  whose elements are taken from an alphabet of  $A$  letters. Nodes represent different sequences and links connect those sequences differing only in one letter. The evolution of a population through the space of genotypes due to mutations is here limited to the NN—or to its largest connected component in case the NN is disconnected. An evolution matrix that models such a dynamical process is

$$\mathbf{M} = f(1 - \mu)\mathbf{I} + \frac{f\mu}{(A-1)l}\mathbf{G}, \quad (3.5)$$

where  $\mathbf{I}$  is the identity matrix and  $\mathbf{G}$  is the adjacency matrix of the connected network, with elements  $G_{ij} = 1$  if nodes  $i$  and  $j$  are connected and  $G_{ij} = 0$  otherwise. The genotypic robustness of a node is proportional to its degree  $k_i$ , defined as the number of genotypes one-mutation away that are on the network,  $k_i = \sum_j G_{ij}$ .  $\mathbf{M}$  describes a population that every time step replicates at each node at a rate  $f > 1$ , each daughter sequence leaving the node with probability  $0 < \mu < 1$  and surviving with probability  $k_i\mu/(A-1)l$  [103], with  $k_i$  the degree of the parental node. If we define  $k_{\min}$ ,  $k_{\max}$  and  $\langle k \rangle$  as the smallest, largest and average degree of that NN, respectively, we obtain  $k_{\min} < \langle k \rangle \leq \gamma_1 < k_{\max}$  for any heterogeneous network,  $\gamma_1$  being the largest eigenvalue of the adjacency matrix  $\mathbf{G}$  [104]. In the case of two-letter alphabets,  $A = 2$ ,  $\gamma_1$  is bounded from above by the logarithm of the number of genotypes in an NN [105].  $\gamma_1$  also equals the average degree of the population at equilibrium,  $\kappa$ , so the former inequality implies  $\kappa > \langle k \rangle$ , indicating that the population

**Box 1.** Dynamics of replicators on a fitness landscape.

The evolution of a population of asexually replicating individuals on a fitness landscape described as a genotype network can be written as

$$\mathbf{n}(t) = \mathbf{M}\mathbf{n}(t-1) = \mathbf{M}^t \mathbf{n}(0) = \sum_{i=1}^m \lambda_i^t (\mathbf{n}(0) \cdot \mathbf{u}_i) \mathbf{u}_i, \quad (3.2)$$

where  $\mathbf{u}_i$  and  $\lambda_i$  are the eigenvectors and eigenvalues of the evolution matrix  $\mathbf{M}$  and  $m$  is the number of nodes of the genotype network;  $\mathbf{n}(t)$  has length  $m$ . We order the eigenvalues and eigenvectors such that  $\lambda_i \geq \lambda_{i+1}$ . If  $\mathbf{M}$  is primitive, Perron–Frobenius theorem for non-negative matrices ensures that, over time, the system evolves towards an asymptotic state characterized by the (unique) first eigenvector  $\mathbf{u}_1$ . More precisely

$$\lim_{t \rightarrow \infty} (\lambda_1^t \alpha_1)^{-1} \mathbf{n}(t) = \mathbf{u}_1, \quad \alpha_1 = \mathbf{n}(0) \cdot \mathbf{u}_1 > 0, \quad (3.3)$$

regardless of the initial condition  $\mathbf{n}(0)$ . The components of  $\mathbf{u}_1$  (all of them guaranteed to be strictly positive by the same theorem) are proportional to the fractions of the total population at each node once the process has reached mutation–selection equilibrium, while its associated eigenvalue  $\lambda_1$  represents the asymptotic growth rate of the population. The transient dynamics towards equilibrium is ruled by the subsequent eigenvalues, but in most cases the time to reach the equilibrium state verifies  $t_{\text{eq}} \propto [\ln(\lambda_1/\lambda_2)]^{-1}$ , since the contributions of higher-order terms are suppressed exponentially fast [103].

In a population of replicators that mutate with probability  $0 < \mu < 1$  per genotype and replication cycle, matrix  $\mathbf{M}$  can be decomposed as<sup>1</sup>

$$\mathbf{M} = (1 - \mu)\mathbf{F} + \frac{\mu}{S}\mathbf{G}\mathbf{F}, \quad (3.4)$$

where  $\mathbf{F}$  is the diagonal matrix  $F_{ij} = f_i \delta_{ij}$ ,  $f_i$  being the fitness (i.e. replication rate) of node  $i$ ;  $\mathbf{G}$  is the adjacency matrix of a connected graph, whose elements are  $G_{ij} = 1$  if nodes  $i$  and  $j$  are connected and  $G_{ij} = 0$  otherwise; and  $S$  stands for the maximum number of neighbours of a genotype [23]. When replicators are sequences of length  $l$  whose elements are taken from an alphabet of  $A$  letters, the size of the genotype space is  $m = A^l$  and  $S = l(A - 1)$ .

Matrices such as  $\mathbf{M}$  in (3.4) are guaranteed to be primitive if the network  $\mathbf{G}$  is connected and the diagonal of  $\mathbf{F}$  is strictly positive.

Dynamics on a single NN is a particular case for which the fitness components are  $f_i = f$  if  $i$  is a genotype in the NN and 0 otherwise—all sequences replicate at a rate  $f$ .

selects regions with connectivity above average on the NN. This fact shows a natural evolution towards mutational robustness, because the most connected nodes are those with the lowest probability of experiencing lethal mutations. Nonetheless, the population might get trapped in regions of lower connectivity if  $N\mu < 1$  [106]. The tendency towards robustness does not preclude evolutionary innovation, though. On the contrary, NNs relevant in evolution span large regions in genome space [50], with the result that they can be more robust and at the same time more evolvable [80,107,108]. A positive correlation between neutrality and evolvability stems from the fact that NNs are very interwoven: for example, all common RNA structures of length  $l$  can be found within a small radius of a randomly chosen sequence in genotype space—a property known as ‘shape space covering’ [84,109]. The mutual proximity of NNs in genome space (the so-called NN apposition [58,110]) has been observed empirically. Two remarkable examples are ribozymes and viruses. Indeed, two RNA sequences with independent origins can fold and function as different ribozymes when their sequences are forced to evolve to increase their similarity, eventually differing in only two nucleotides [42]; diffusion on NNs is instrumental to permit innovation and immune escape in influenza A [24].

The eigenvectors of the adjacency matrix  $\mathbf{G}$  are also eigenvectors of the evolution matrix  $\mathbf{M}$ , as can be seen in equation (3.5). Their respective eigenvalues,  $\gamma_i$  and  $\lambda_i$ , are different—albeit related through  $\lambda_i = f(1 - \mu) + \gamma_i f \mu / (A - 1)$ . As a consequence, in NNs the asymptotic state of the system only

depends on the topology of the NN, and parameters such as the mutation rate  $\mu$  or the sequence length  $l$  exclusively affect the transient dynamics towards equilibrium [103,106]. This result cannot be extrapolated to more general fitness landscapes, where both the equilibrium state of the population and the transient dynamics depend in a non-trivial fashion on network topology and genotype fitness [103] (cf. equations (3.2) and (3.4) in box 1).

Heterogeneity in the degree of the nodes, or equivalently in genotypic robustness, and the assortativity inherent to many NNs have important consequences in the dynamics of populations. Soon after the hypothesis of the molecular clock [111] was put forward, variations in genotypic robustness were suggested as an explanation for its unexpected overdispersion [112]. If networks are furthermore assortative, the probability that the population leaves the network diminishes the longer the time spent on it, leading to a progressive (phenotypic) entrapment. Beyond a systematic increase in the overdispersion of the process with time, assortativity entails an acceleration in the fixation rate of neutral mutations [79], invalidating the Poissonian assumption underlying the molecular clock.

## 4. Punctuated dynamics in molecular adaptation

As soon as more realistic architectures of the genotype space are considered, dynamics becomes punctuated. This fact has

**Box 2.** Competition for centrality in a network of networks.

In complex network theory, the eigenvector centrality  $x_k$  of a node  $k$  in a network is defined as the  $k$ th component of the eigenvector of its adjacency matrix  $\mathbf{G}$  corresponding to the largest eigenvalue  $\gamma_1$  [116]. The eigenvector centrality has become one of the most widespread metrics for node importance because of its wide range of applications, which include Google Pagerank [117], estimations of the professional impact of scientists [118] and journals [119], the importance of individuals in a social group [120] or of regions in the brain [121], and dynamical processes such as disease or rumour spreading (see [116] for an overview).

This measure can be generalized to other dynamical processes if  $\mathbf{G}$  is replaced by another (non-negative) matrix  $\mathbf{M}$ : the new eigenvector centrality is defined through  $u_1$ , the eigenvector corresponding to  $\lambda_1$ , the largest eigenvalue of  $\mathbf{M}$  (see e.g. box 1). In evolutionary dynamics, the eigenvector centrality is thus the fraction of population with each genotype at mutation-selection equilibrium [103]. We use this generalization in the following.

In a network of networks, the centrality of each network is the sum of the centralities of all its nodes, normalized in such a way that the sum of the centralities of all networks is equal to one. Therefore, combining game theory and network science, we can approach the spread of the total centrality on the different networks as a zero-sum game, where players are not nodes but networks and *compete* for centrality, which is understood as a limited resource. The winnings of each competing network  $\alpha$  are calculated as the total centrality  $C_\alpha$  accumulated by all its nodes

$$C_\alpha = \frac{\sum_{j \in \alpha} u_{1j}}{\sum_{k=1}^m u_{1k}},$$

where  $j$  runs on the nodes of network  $\alpha$  and  $m = \sum_{\mu} m_{\mu}$  is the total number of nodes in the network of networks. The outcome of such confrontations for centrality and the time needed by the winner to prevail drastically depend on (i) the internal structure of the competing networks  $\alpha = 1, \dots, K$ , as characterized by their maximum eigenvalue  $\lambda_{1,\alpha}$  in a way that networks with larger  $\lambda_{1,\alpha}$  in general obtain more centrality than their competitors and (ii) the connector nodes, that is, the boundary nodes that connect one of these networks with the rest of them through connector links.

When connector links occur only through nodes with little centrality (aka peripheral connections), almost all centrality remains in the network with the largest eigenvalue  $\lambda_{1,\alpha}$ . If for some reason (e.g. an environmental change) the eigenvalue of a different network overcomes  $\lambda_{1,\alpha}$  a sharp centrality redistribution takes place. The time to reach the equilibrium significantly increases close to that transition.

been highlighted in formal studies stating that GP maps based on RNA sequence-to-structure relationship naturally imply punctuation, irreversibility and modularity in phenotype evolution [21], and has been nicely illustrated in computational works [22,58,110].

The formal scenario that we use here starts at the level of genotypes, but also takes into account the non-trivial topology induced by the mapping onto phenotypes. By means of techniques that exploit the networked and modular structure of genotype spaces, we will show that the dynamical behaviour is qualitatively similar in three different situations, that is if (i) an NN has two or more regions of high connectivity linked through few possible mutational pathways, (ii) a population encounters a phenotype of fitness higher than the extant one or (iii) mutation-selection equilibrium is perturbed through an environmental change that entails a modification of the fitness landscape. Underneath the punctuated dynamics observed in those situations there is a common mechanism: a (formal) competition between regions with a high internal connectivity that are sparsely connected to one another. These highly internally connected regions may be different clusters of genotypes in a single NN, different phenotypes each characterized by its own NN, or different regions in a fitness landscape. Actually, this synthesis emerges as a generalization of processes occurring on a wide variety of biological, technological and social dynamics on networks of networks (i.e. networks connected through a limited number of connector links). This class of processes admits a description in terms of competitive scenarios where each network is

defined as an independent agent struggling with the rest for a particular kind of resource [113–115]: *eigenvector centrality* (see box 2).

#### 4.1. Metastable states and punctuation in a network-of-networks architecture

In §3, we have focused on the dynamics of populations evolving on a single NN characterized by a well-defined region of maximum connectivity. Under those conditions, the evolutionary dynamics of a sufficiently large population is smoothly canalized towards the maximally connected region of the NN [83,103,106]—something that has measurable effects on the fixation rate of neutral mutations [79]. However, there is no *a priori* reason to assume that generic NNs do not present a complex structure formed by more than one cluster of nodes with high internal connectivity and sparse connections to one another. For instance, the topology of NNs associated with RNA secondary structures recently revealed an intricate network-of-networks organization, where the different communities can be further divided into subcommunities attending to their sequence composition [122, Fig. 6]. Some of these networked communities turned out to be two mutational steps away—such mutations playing the role of what we have named connector links—and therefore required an intermediate group of genotypes for a population to move from one community to another. In this type of complex structure, the evolutionary dynamics of populations on NNs can display an alternance

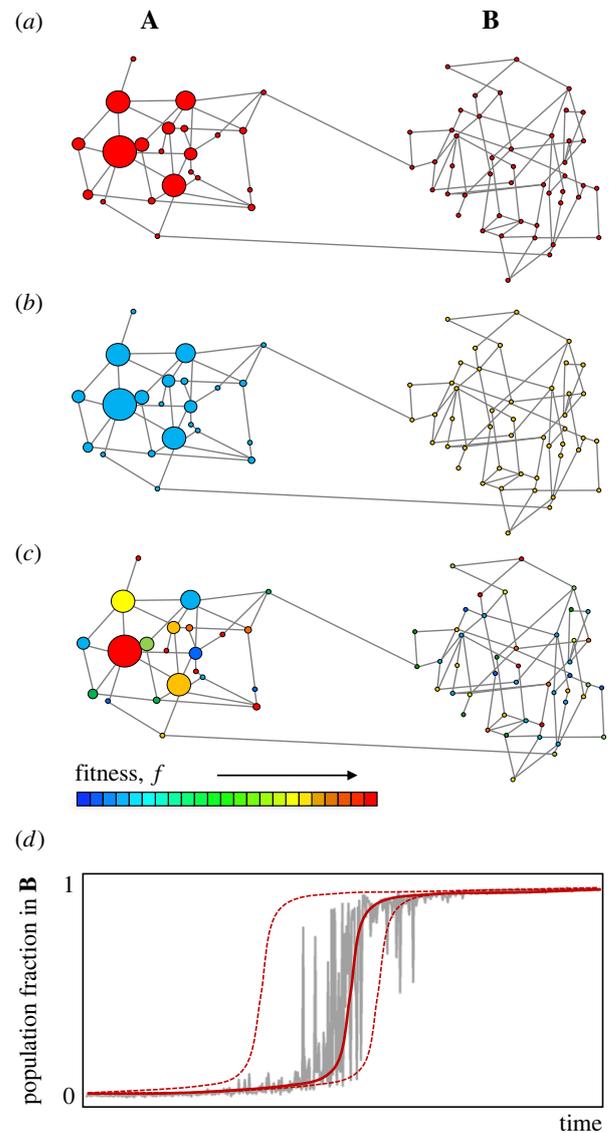
of metastable states (which might appear as true equilibria at short times) with periods where neutral mutations are rapidly fixed [102].

The formalism that describes competition between networks for centrality, while originally introduced in the framework of complex network theory, was recently proven to be fully applicable to the study of populations evolving in the space of genotypes [101]. The population distribution at mutation-selection equilibrium is given by the first eigenvector  $\mu_1$  of the matrix  $\mathbf{M}$  that characterizes the dynamical process, and therefore the centrality that each network competes for coincides with the fraction of organisms that populate its corresponding sequences in the asymptotic state. In general, the most populated network in the equilibrium is the one with the largest eigenvalue  $\lambda_1$  of matrix  $\mathbf{M}$  (box 2).

Let us illustrate in the simplest case how a population moves from a subnetwork with a lower eigenvalue  $\lambda_{1,A}$  to a subnetwork with a larger eigenvalue  $\lambda_{1,B}$  in the framework of competition for centrality. Figure 3*a* represents two regions of an NN weakly connected. As previously described, we have  $\lambda_{1,A} = f(1 - \mu) + \gamma_A f \mu / (A - 1)l$ , and similarly for network **B**. Note that the latter network will eventually be attracting the population if the eigenvalue corresponding to its evolution matrix  $\lambda_{1,B}$  is larger than that of **A** and, as a consequence, the same applies to the adjacency matrices (i.e.  $\gamma_B > \gamma_A$ ). This result shows that the separating barrier only depends on the topological structure (size and connectivity) of each subnetwork. The transition to a region with higher connectivity occurs upon stochastic appearance of mutations along connecting pathways. This process is highly contingent, so the time of the punctuation is difficult to predict (red lines in figure 3*d*). Actually, too small populations might be indefinitely trapped in regions as **A** [22].

## 4.2. Drift and switch dynamics in adaptive transients

Early evidences of punctuation in molecular adaptation came from computational simulations of populations of RNA sequences evolving towards a target secondary structure [22]. Typically, populations remain on the current phenotype until a higher-fitness solution is found, that is, until one of the genotypes in the population acquires a mutation that produces a new, fitter phenotype. This event is preceded by a ‘search’ in the original phenotype during which the population accumulates neutral mutations and increases its genotypic diversity. The switch transition is not deterministic, since different phenotypes can be reached first depending on the stochastic occurrence of mutations. Once the new phenotype has been found, the transition occurs exponentially fast but, concomitantly, the population experiences a severe bottleneck that reduces its genotypic diversity. In this scenario, a new phenotype can be accessed through any genotype in the neighbourhood of genotypes of the original phenotype, though peripheral genotypes (those with a higher number of links pointing to different phenotypes, i.e. of low robustness) are more likely to act as connectors than highly robust, central genotypes [79]. This drift and switch dynamics is characteristic of any realistic GP map with a structure such as that described in §2. In the dynamical framework of competition between networks, each phenotype represents now a distinguishable network characterized by its size, connectivity and fitness level. Connector links correspond to regions of apposition between the two networks, which exist in most cases (in



**Figure 3.** Genomic shifts result from the network-of-networks structure of the space of genotypes. Without loss of generality, we assume that  $\lambda_{1,A} < \lambda_{1,B}$  and the whole population is initially in network **A**. In (a–c), colours indicate the fitness of each node, as shown by the colour scale, and circle size is indicative of the number of individuals at each node. Though nodes in network **B** are represented with small circles, we assume they have no population initially. (a) Two weakly coupled regions of a unique NN. Differences in their eigenvalues only depend on differences in their topology. (b) Two different NNs with different fitness. The effect of fitness and topology can be separated, both affect their eigenvalues. (c) Two weakly connected regions in a fitness landscape. The effects of fitness and topology cannot be decoupled. (d) In all cases, the time of transitions is a stochastic variable, but the transition is fast once the mutational pathway is found (red curves, corresponding to different realizations of the process). In changing or noisy environments, the fitness value of each sequence might vary in time, so transitions are decorated by fluctuations (grey curve) whose strength grows as the tipping point is approached.

particular when the two phenotypes considered are common) but are difficult to find if populations are finite due to the vastness of genotype spaces and NN [30]. Also, the connector links might join regions with similar fitness but different internal connectivity, or regions with different fitness, among many other possibilities. Different paths to adaptive improvement are taken with different probability. For example, narrow neutral paths are crossed much faster than fitness valleys [123].

Figure 3*b* illustrates the situation of two phenotypes with different fitness values (i.e. replicative ability of its nodes) coupled through narrow paths. The transition to phenotype B might occur if  $\lambda_{B,1} > \lambda_{A,1}$  which implies that

$$\frac{f_A}{f_B} < \frac{1 - \mu + \mu\gamma_B/(A-1)l}{1 - \mu + \mu\gamma_A/(A-1)l} \approx 1 + \frac{\mu}{(A-1)l}(\gamma_B - \gamma_A), \quad \mu \ll 1, \quad (4.1)$$

where the specific effect of fitness  $f_i$  and topology  $\gamma_i$  is quantified.

The survival-of-the-flattest effect represents one particular case of such competition where the two competing regions have different levels of fitness, different mutation rates (a situation that can be easily included in the framework above), and different levels of robustness [124,125], which effectively accounts for different topologies [126]. Some computational models that consider relevant features of molecular populations have been developed, leading to an improved understanding of this interesting phenomenon. The effective implementation of networks with different degrees of neutrality permits to capture competitions between RNA populations subject to selection for different folds (characterized by different neutral networks), showing how the relative advantage of either population changes sign as a function of the mutation rate [127]. Also, in a model of quasi-species characterized by bit strings, it has been shown that a discontinuous phase transition separates the regions of selection for replication and selection for robustness [128]. At odds with results emphasizing selection for robustness along evolution, models with RNA quasi-species show that high mutation rates might be evolutionarily advantageous in situations where a single RNA sequence might code for a molecular ecosystem [129]. It has been also argued that fitness landscapes with fitter and flatter regions might be behind the enigmatic richness of microbial metabolisms [130]. Epochal evolution (i.e. metastable states punctuated by rapid transitions to fitter states) have also been observed in evolutionary search algorithms, as referred to a class of optimization techniques [131,132].

The theory can be easily extended to any number of phenotypes in competition and yields a clear prediction regarding the phenotype that will be eventually attracting the population. The largest eigenvalue of any matrix  $\mathbf{M}$ ,  $\lambda_1$ , is a fundamental quantity that synthesizes information on the topology of the underlying network, on the fitness of its nodes and on the mutation rate. These three elements combine in a non-trivial way to determine the competitive ability of a population on a given network. In this respect, a population can asymptotically displace a competitor for a number of different reasons, namely because (i) it spreads on a larger NN, (ii) its average fitness is higher, (iii) it spreads on a network with higher connectivity, (iv) it mutates at an advantageous rate with respect to its competitors or (v) any suitable combination of the previous reasons.

### 4.3. Smooth environmental changes and genomic shifts

There is empirical evidence that environmental changes affect the evolutionary dynamics of populations and their eventual fate [133]. Recalling that fitness is an environment-dependent quantity, environmental changes can be formally cast as modifications of the fitness associated with genotypes. When a genotype space is mapped to a realistic fitness

landscape, smooth environmental changes can be represented as gradual modifications of the fitness value of each genotype. As phenotype is here a hidden variable, at this point we do not need to consider possible changes in phenotypic expression due to environmental variation. This possibility will be discussed later though.

Even if environmental variations are smooth, populations may eventually suffer sudden transitions in their genomic composition [23]. In the case of finite populations, there is a non-zero probability of extinction if the pathway linking the (decreasingly fit) current state of the population to a new region populated by fitter phenotypes is not found sufficiently fast [101]. The abundance and breadth of connecting pathways depends on the roughness of the landscape and on the fraction of lethal mutations, which can be put in correspondence with important variables such as the degree of heterogeneity of the corresponding genotype networks and the holeyness of the landscape [134]. These quantities tune the number of connector links between different regions with significant fitness and the centrality of their connector nodes. As a consequence of the above, fitness landscapes can be described as a network of networks formally analogous to the examples discussed previously (figure 3*c*).

Early warning signals that forecast the proximity of tipping points (and therefore of a putative extinction threshold) can be defined in analogy to studies of sudden shifts in ecology [14]. Close to those state transitions populations show flickering and hysteresis, i.e. a dependence on its previous states that causes trapping and metastability, and is eventually responsible for extinction [101].

Summarizing, facing evolutionary systems from the viewpoint of competing networks turns the space of genotypes into a network of networks at several different levels. The full consequences of this architecture are still to be understood, though they are certainly far from trivial: relevant phenomena such as robustness [135,136], synchronization [137,138], cooperation [115,139,140] or epidemic spreading [141–143] exhibit different features when their dynamics occur on a single network or on a network of networks.

## 5. The many-to-many nature of the genotype – phenotype map

Our discussion so far has assumed that each genotype corresponds to a unique phenotype. Adaptation to a new environment or selection pressure, therefore, has to be achieved through mutations, and we have discussed some of the non-trivial phenomena that appear when heterogeneous populations evolve in a complex genotype space. However, there are many cases in which genotypes express more than one phenotype, opening up new possibilities for adaptation: in any realistic realization, the GP map is many-to-many, since genotypes are able to express different phenotypes in a variety of situations. In this section, we present several examples of this phenomenon and discuss how it alters the dynamics discussed in previous sections. The reader should know that the level of formal description achieved is poorer than for dynamics on networks and has received much less attention up to now. Our feeling is that, as shown in previous sections, theory should help towards unifying processes and concepts that are treated at present as different phenomena. However, the following sections rely much more on the description of the

latter than on quantitative results. A full mathematical formalism that describes at once the multilayered, network-of-networks structure of the genotype-to-function map is an open and on-going problem of the highest relevance.

### 5.1. Molecular promiscuity

Enzymes were classically thought to be highly specific: one enzyme–one substrate–one reaction. However, recent experimental data have shown that, in fact, many enzymes are able to catalyse more than one reaction, a phenomenon that has been termed catalytic or functional promiscuity [144–149]. This means one amino acid sequence corresponds to more than one phenotype. Promiscuous enzymes are not hard to find in sequence space. For example, single-site mutants of bacterial enolases can actually perform secondary functions not found in the wild-type, while maintaining their original activity [150]. Moreover, these promiscuous functions are easily evolvable: enzymes can accumulate mutations that do not alter their main function, but which radically change their secondary ones [151–153], and the activity of secondary functions can be increased several orders of magnitude with very few mutations [144,154,155].

Promiscuous activities can help enzymes evolve towards new functions. A polymorphic population of enzymes can diversify with respect to its secondary functions if they bear no fitness costs to the organism, leading to the accumulation of what has been termed cryptic genetic variation [156]. When selection pressure for a new function appears, those enzymes in the population that carry out that function as a promiscuous activity will be already functional and, in a sense, pre-adapted for it. The new function can then be improved through over-expression [147] or gene duplication that liberates one copy of the enzyme to specialize in the new function [144,148,157]. These promiscuous activities also have an effect on metabolism, connecting different metabolic pathways [148,158], and therefore enabling their gradual evolution: promiscuous enzymes can develop their secondary functions, so that certain steps in a pathway become more efficient, in turn liberating other enzymes to focus on other parts of the pathway. The evolution of metabolic pathways, therefore, can be achieved in a more parsimonious way. When a new pathway is needed, cells with promiscuous enzymes may perform the needed reactions, and give these sequences an adaptive advantage.

Functional promiscuity is not restricted to enzymes: transcription factors have been shown to bind many different motifs with comparable binding energies [32,44,148]. Also, proteins can be mistranslated [159], a process that is several orders of magnitude more common than genetic mutations, and thus at a given moment in time, some proteins will have a different amino acid sequence, with potentially different functions that can accelerate adaptation to a new function [160–162]. Some protein sequences will be more likely to yield new functions under these phenotypic mutations.

Promiscuity is also not restricted to proteins. Early computational work on RNA secondary structures [38] already suggested that RNA molecules could fold into more than one structure, and recent experimental studies have found evidence of RNA molecules that can perform more than one different function [163,164]. The best examples are ribozymes (RNA enzymes) that are able to catalyse two different reactions [42,165,166]. Computational [83,167] and experimental studies

[166] suggest that secondary functions in RNA molecules can evolve as easily as in proteins, and that this functional promiscuity can spread through populations as cryptic genetic variation, accelerating the rate at which new functions are found in evolution. Even if these functions are performed marginally at first, they will give the sequence an advantage if they are selected for, and freedom to improve the new function in genotype space. In fact, theoretical models predict that promiscuous functions can help accelerate evolution towards a new function, through what has been called the look-ahead effect [160]. Although this phenomenon was originally proposed for phenotypic mutations, it is also valid for promiscuous enzymes and RNA molecules.

### 5.2. Phenotypic heterogeneity and bet-hedging

The fact that one sequence can perform more than one function is not restricted to the molecular level. At the regulatory level, for instance, expression noise is very common [168–170], due to the stochastic nature of transcription and translation and the small number of molecules involved in these processes. Expression noise leads to phenotypic heterogeneity [171,172], where two genetically identical genotypes can, under the same conditions, express two different phenotypes at the cellular level. Although expression noise is inherent to the biochemical process of building the phenotype from the genotype, cells can control it to some level [169,173–175], and they can also use it to their advantage [171,172,176]. For instance, genotypes can evolve a stochastic switching mechanism that enables them to alternate between two different phenotypes, a phenomenon that has been termed bet-hedging [177]. At a given moment in time, a fraction of the population will express one phenotype and the rest another one. Each phenotype is typically advantageous in one environment and disadvantageous in another, and so the ability to switch between them is adaptive under some conditions [178]. Typical examples of bet-hedging are bacterial competence [179] and persistence [180]. Bet-hedging is a common mechanism that can also emerge in evolution experiments [181]. These strategies would not be possible without functional promiscuity.

### 5.3. Phenotypic plasticity

Another piece of this puzzle comes from phenotypic plasticity, a well-known phenomenon in which a genotype is able to express different phenotypes in different environments [182]. Notice the difference from phenotypic heterogeneity as discussed above: phenotypic plasticity is only unveiled when an environmental cue appears. In fact, strategies such as bet-hedging arise when the cost of developing a plastic response—which is able to sense the environment—is so high that it becomes disadvantageous [178].

Phenotypic plasticity has been known for a long time in multicellular organisms, but it appears at the unicellular and molecular level as well. Proteins are not only promiscuous: they can also carry out different functions in different environments, a phenomenon that is called moonlighting [183,184]. One classical example is crystallin lenses, enzymatic proteins whose function becomes structural when expressed at very high concentrations [185]. The same gene can also express different proteins through alternative splicing [184]. RNA molecules can fold into different structures at different

temperatures, performing different functions [186]. RNA thermometers, as they are called, can be designed computationally [187]. Gene regulatory networks have different spatio-temporal expression patterns when exposed to different environmental inputs [188–190], and metabolic systems are able to survive on different food sources [68–70].

A plastic population will be able to automatically survive in a new environment, if it expresses a viable phenotype. Once in the new environment, it might spread through the new fitness landscape, maybe losing its original plasticity. Many theoretical and computational studies of plasticity and its relationship with adaptation have been proposed [191–197], although most of them do not include the complexities of the GP map that we have discussed in our previous sections. They assume that phenotypes that are close in trait value to the ones present in the population will always be achievable through mutations. Therefore, the discussion of when and how phenotypic plasticity will be promoted cannot account for the biases induced by more or less abundant phenotypes, asymmetric connections between them and other factors discussed so far in this review, which could affect how easily plasticity is developed. There are, however, some computational studies that explicitly model GP maps, focusing on RNA molecules [83] and gene regulatory networks [189,198].

## 6. Hints for a dynamical theory of many-to-many genotype–phenotype maps

### 6.1. Promiscuity redefines the fitness landscape

How do we integrate all of these data into the framework we have been discussing so far in this review? The presence of phenotypic noise or functional promiscuity (at the molecular or regulatory level) implies that a single genotype, in a given environment, will express more than one phenotype in a probabilistic manner. Therefore, the effective fitness of the genotype will be an intermediate value related to the fitness associated with each phenotype. Naively, one could guess that the fitness  $f_i$  of sequence  $i$  would be  $f_i = \sum_{p \in \mathcal{P}} f(p) \pi_i(p)$ , where  $\mathcal{P}$  is the set of all phenotypes,  $f(p)$  is the fitness of phenotype  $p$  and  $\pi_i(p)$  is the probability that sequence  $i$  expresses phenotype  $p$  (alternatively,  $\pi_i(p)$  represents the fraction of the homogeneous population with genotype  $i$  expressing phenotype  $p$ ). To illustrate one such case, consider a population of RNA sequences that perform their function by interacting with a ligand. Under the minimum free energy mapping usually considered in the literature, all RNA sequences expressing the optimal structure as their minimum free energy are assigned the same fitness. Including promiscuity, however, alters this fitness function. Two sequences belonging to the same NN have different compositions, and this variation leads, in general, to differences in their folding energies and also in the repertoire of structures with which they are compatible [199]. Differences in the folding energy entail differences in the average time spent in the minimum free energy secondary structure for each specific sequence. In this situation, a more accurate definition of fitness takes it as proportional to the time spent in the optimal secondary structure. Therefore, two sequences belonging to the same NN have different fitness values under this more realistic quantification of their function.

However, a careful investigation of the underlying (stochastic) population dynamics reveals that the simple average above is not of general applicability, as the next example illustrates. Consider a homogeneous population of cells expressing a certain phenotype with probability  $p$ , and another one with probability  $1 - p$ . The replication rate  $\beta$  of both phenotypes is the same, but the second phenotype has a higher death rate  $\delta_2 > \delta_1$ —i.e. it has a lower fitness, defined as the difference between birth and death rates,  $f = \beta - \delta$ . There is no mutation in this example. Whenever any cell replicates, the daughter cell expresses one of the two phenotypes with the aforementioned probabilities, regardless of the mother's phenotype. Calling  $m_1(t)$  and  $m_2(t)$  the number of cells of each type at time  $t$ , we can use results from birth–death processes theory to derive the following system of ordinary differential equations:

$$\begin{pmatrix} \dot{m}_1(t) \\ \dot{m}_2(t) \end{pmatrix} = \begin{pmatrix} \beta p - \delta_1 & \beta p \\ \beta(1-p) & \beta(1-p) - \delta_2 \end{pmatrix} \begin{pmatrix} m_1(t) \\ m_2(t) \end{pmatrix}. \quad (6.1)$$

We diagonalize the system to obtain its largest eigenvalue (and thus, the asymptotic fitness of the population):

$$\lambda_1 = \frac{\beta - \delta_1 - \delta_2}{2} + \sqrt{\frac{(\beta - (\delta_2 - \delta_1))^2}{4} + \beta p (\delta_2 - \delta_1)}. \quad (6.2)$$

With some algebra, we can show that  $\lambda_1 > (\beta - \delta_1)p + (\beta - \delta_2)(1 - p)$ , the latter being the result of the naive guess above, i.e. that the average fitness of the population is the weighted average of the fitness of the visited phenotypes, where weights are the probability that a genotype expresses each phenotype. The discrepancy arises, in this case, because cells expressing the second phenotype die more often. As a result, the population has an overrepresentation of cells expressing the more stable phenotype: their fraction in the population is actually greater than  $p$ .

Despite the differences between the two examples discussed, it appears that the effect of promiscuity can be accounted for by properly redefining the fitness landscape. Each example, however, will need to be carefully examined to correctly translate its dynamical details to a suitable definition of fitness.

### 6.2. Dynamics of plastic phenotypes under frequent environmental changes

Phenotypic plasticity means that the same genotype expresses different phenotypes in different environments, such that different evolution matrices have to be considered in each of the environments (see box 3). This is equivalent to considering one GP map per environment, and switching between them when the environment changes. To fix ideas, suppose we have two different environments alternating every generation, with associated matrices  $\mathbf{M}_1$  and  $\mathbf{M}_2$ . Then the evolution of the population will be given by the largest eigenvalue of the matrix  $\mathbf{M}_2 \mathbf{M}_1$  and the asymptotic state of the population turns out to be an orbit with period 2, as long as some conditions are fulfilled. Both matrices (and their product) must be primitive (see box 1). This happens, for instance, if all nodes have positive fitness or if, after removal of the zero-fitness nodes, none of the two networks breaks down into different connected components. If this condition is not met, the asymptotic state will depend on the initial condition. Likewise, even if all nodes have positive

**Box 3.** Dynamics of replicators on a shifting fitness landscape.

The framework introduced in box 1 can be extended to account for environmental changes. For the sake of simplicity, we will just consider the case in which the environment alternates between two states, but generalizations of this are self-evident. The fitness of every node needs not be the same in each environment, and as a result the evolution matrices of both environments (we will denote them by  $\mathbf{M}_1$  and  $\mathbf{M}_2$ ) will be different.

Let us begin by exploring the case in which, starting in environment 1, we alternate environments every generation. Then the equation for the evolution of the population reads

$$\mathbf{n}(t) = \begin{cases} [\mathbf{M}_2\mathbf{M}_1]^{t/2}\mathbf{n}(0), & t \text{ even,} \\ [\mathbf{M}_1\mathbf{M}_2]^{(t-1)/2}\mathbf{M}_1\mathbf{n}(0), & t \text{ odd.} \end{cases} \quad (6.3)$$

This means that, in general, the evolution of the population will be dominated by the largest eigenvalue of the matrix  $\mathbf{M}_2\mathbf{M}_1$  at even times and of the matrix  $\mathbf{M}_1\mathbf{M}_2$  at odd times, regardless of  $\mathbf{n}(0)$ . (Starting from environment 2 would only swap the parity of times, but not the general results.)

Interestingly, the eigenvalues of cyclic permutations of a product of matrices are the same, and the corresponding eigenvectors are easily related to each other. Thus, if  $\lambda_1$  is the largest eigenvalue of  $\mathbf{M}_2\mathbf{M}_1$  and  $\mathbf{v}_1$  its corresponding eigenvector, then the eigenvector of matrix  $\mathbf{M}_1\mathbf{M}_2$  will be  $\mathbf{M}_1\mathbf{v}_1$ , so the asymptotic population will grow as  $\lambda_1^{t/2}$  and the fraction of population will cycle through

$$\mathbf{v}_1 \rightarrow \frac{\mathbf{M}_1\mathbf{v}_1}{|\mathbf{M}_1\mathbf{v}_1|} \rightarrow \mathbf{v}_1. \quad (6.4)$$

The case in which environments change following a random pattern is particularly interesting. In this case,

$$\mathbf{n}(t) = \mathbf{M}^t\mathbf{n}(0), \quad \mathbf{M} \equiv \left\langle \prod_{k=1}^t \mathbf{M}_{\mu_k} \right\rangle^{1/t}, \quad (6.5)$$

where  $\mu_k \in \{1, 2\}$  is a discrete random process whose dynamics is prescribed (for instance, it can take each of the two values with a certain probability, or  $\mu_1$  can take any value with a certain probability and swap every time step with another probability). The expected value is to be taken over realizations of this process. The largest eigenvalue of  $\mathbf{M}$  and its corresponding eigenvector will determine the asymptotic behaviour of the population. Mathematically, this process is not fully characterized yet, but it is not difficult to carry out its numerical implementation.

fitness but the fitness of some of them is very small, the population can get trapped in metastable states for very long times. But one can also imagine that alternating environments can have the opposite effect, namely, that the transit of certain pathways strongly hindered in both environments when kept constant may be facilitated by their alternation.

This analysis can be extended to more complicated alternating patterns of the two environments, the only difference being that the asymptotic state will exhibit a longer period. For instance, if environments change according to the pattern 112112112... , and  $\lambda_1$  and  $\mathbf{v}_1$  are the largest eigenvalue and its corresponding eigenvector of the matrix  $\mathbf{M}_2\mathbf{M}_1^2$ , then the population will grow as  $\lambda_1^{t/3}$  and the fraction of population will cycle through

$$\mathbf{v}_1 \rightarrow \frac{\mathbf{M}_1\mathbf{v}_1}{|\mathbf{M}_1\mathbf{v}_1|} \rightarrow \frac{\mathbf{M}_1^2\mathbf{v}_1}{|\mathbf{M}_1^2\mathbf{v}_1|} \rightarrow \mathbf{v}_1.$$

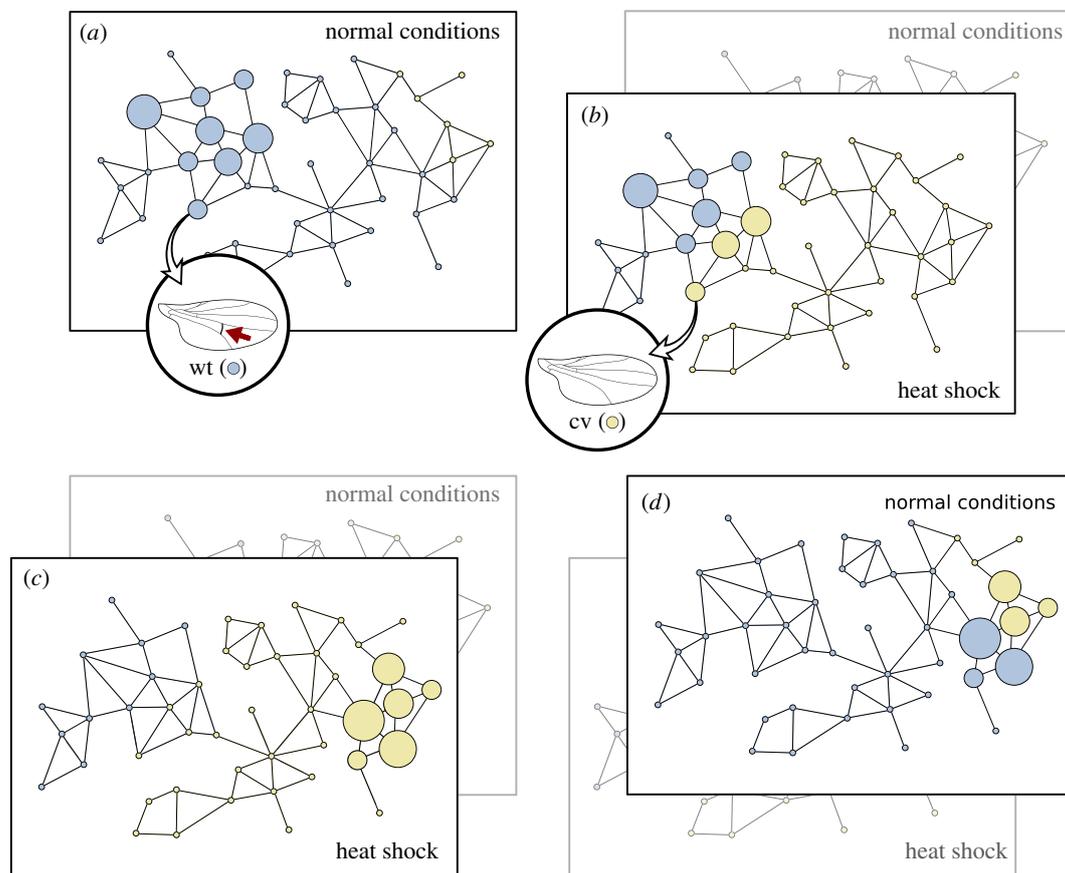
A qualitative representation of this idea was already proposed in the form of adaptive multiscapes [30] (figure 4). It was shown there that the evolutionary phenomena introduced by phenotypic plasticity, such as Waddington's genetic assimilation [34], could be easily understood in terms of a multilayered network of genotype networks. Genetic assimilation is a very interesting phenomenon. In Waddington's experiment, a plastic population of flies was exposed to a new environment, in which they expressed a different phenotype (called cross-veinless). They were selected for this new phenotype under the new environment, so they spread through the genotype network in the way we have discussed

in §4. After some time, when the population was brought back to the original environment, some of the individuals kept the cross-veinless phenotype, instead of reverting to the wild-type (figure 4). The phenotype that originally appeared only plastically was now being expressed without environmental changes: it had become genetically assimilated. Adaptive multiscapes help in the qualitative understanding of the molecular mechanisms underlying genetic assimilation, among others, since the population dynamics sketched in box 3 suffice to explain it.

## 7. Discussion and prospects

A large body of current evidence shows that the gradualistic view of evolution is at odds with the mechanisms operating at the molecular level, where discontinuous changes and fast pre-adaptations are the rule rather than the exception. We have presented three basic mechanisms with a strong effect on the evolutionary dynamics of biomolecules: fast exploration of new phenotypes by heterogeneous populations spread over neutral networks, competition between different networks for population (the evolutionary counterpart of eigenvalue centrality) and plasticity of phenotypes. But ubiquitous and general as they may be, these are by no means the only ones. Several other mechanisms and phenomena have been left out from our framework.

The first one has to do with mutations. The most parsimonious change in a genome is represented by point mutations. All through this review, we have shown how even these



**Figure 4.** Waddington's genetic assimilation under the light of genotype networks. Each layer of the network represents a different environment. Here, there are two environments: normal conditions and heat shock. As in previous figures, circle size is proportional to the number of individuals populating that node—small circles represent unpopulated nodes. The colour of each node represents now its phenotype, instead of its fitness. Note that every genotype appears in both layers, and that connections between them are the same in both environments: the only property that changes is the phenotype. (a) A population of flies develops wings with a cross-vein (the wild-type phenotype, wt, blue) when bred in normal conditions. (b) When exposed to heat shock during development, some of the flies in the original population develop new wings without cross-veins (the cross-veinless phenotype, cv, yellow). (c) Breeding the flies under heat shock and then selecting for those flies expressing the cross-veinless phenotype, the population drifts towards a new part of genotype space, exploring a new neutral network (or possibly increasing fitness in the new environment). (d) After some time, the population is bred again in normal conditions, and some flies in the population keep expressing the cross-veinless phenotype. Their phenotype has been genetically assimilated.

minor changes frequently cause major phenotypic modifications. The evolution of genomes, however, is often driven by mutational mechanisms that substantially modify them, such as gene duplication or horizontal gene transfer (HGT). The latter will potentially cause effects of magnitude larger than point mutations, and therefore entail still stronger effects on phenotypes and functions. The structure of genomes, especially the existence of universal regularities in the distribution of genomic elements [200], speaks about dominant mechanisms beyond organismal adaptation [201,202]. Gene sharing through HGT has played a main role in the adaptation of microorganisms [203] and is so common in microbial evolution that it has led to the idea of network genomics [204]. The reconstruction of gene-sharing networks for viruses [205] has uncovered a hierarchical and modular structure that drastically changes our view of viral species as well-defined entities. Instead, the topology of such networks reveals an utmost plastic system where genes behave as highly mobile pieces, and where not only adaptation but also evolutionary innovations might be strongly promoted through combinatorial processes—especially in viruses with segmented genomes [206]. This plastic view of the genome can be straight forwardly extended to cellular organisms.

Secondly, we have not included any kind of sexual reproduction nor recombination—of which HGT is a particular

case. Though recombination might slow-down evolution under strong selection [207], in most of its forms it is a powerful enhancer of the search for novelty [208]. This power is very well illustrated in experiments of DNA shuffling [209], where a chimaeric cephalosporin created from recombination of four different ones achieves a 270-fold increase of resistance to antibiotic—compared to the eightfold increase achieved by the best cephalosporin created through point mutations alone. On top of that, the interplay between recombination and the genotype–phenotype map may induce a fascinating disruptive dynamics that resembles sympatric speciation [210], so speciation—one of evolution's major themes—may not be properly understood unless recombination is suitably incorporated in our dynamical models. However, this cannot be done if size- and frequency-dependent evolution operators are not introduced, because the probability that a recombination event takes place depends on the relative presence in the population of the sequences to be recombined. The lack of a suitable framework to describe this complication leaves any 'ecological' interaction between molecules or genes out of the picture. This is probably the weakest point of the network formalism—one that is of paramount importance to tackle in future work.

Even if we constrain ourselves to the range of applications to which the formalism we are advocating does apply, its

actual implementation is not free from serious difficulties. To begin with, the vastness of genotype spaces makes it impossible to explore any realistic genotype–phenotype map in depth. This is a handicap that will not be solved with more powerful computers, so we need to turn to an alternative description of evolutionary dynamics. Fortunately, all models of the genotype–phenotype map share a set of common properties regardless of the details. This situation is similar to the one faced by Statistical Physics in its aim to go from microscopic models to macroscopic description, and so it can be dealt with in a similar vein. If details do not matter, we may try to build a mesoscopic description in which phenotypes, rather than genotypes, are the basic elements of our dynamical framework, and in which microscopic details are subsumed in an effective, possibly non-Markovian stochastic dynamics [79].

We also need to figure out how to incorporate promiscuity and environment in our evolutionary picture, in a way that does not require the running of specific simulations for each particular case. If a mesoscopic description is to be made, any change in the environment would entail a full reconfiguration of the network of phenotypes, thus affecting not only the phenotype that the population currently occupies but also the transitions between different

phenotypes—hence the evolutionary pathways. A way to incorporate the effect of the environment would be through a multilayer formalism for networks [85,86], where different layers would correspond to different environments. Generalizing the dynamics described here to a multilayer network is as yet an open problem.

**Data accessibility.** This article has no additional data.

**Competing interests.** We declare we have no competing interests.

**Funding.** This work has been supported by the Spanish Ministerio de Economía y Competitividad and FEDER funds of the EU through grants ViralESS (FIS2014-57686-P) and VARIANCE (FIS2015-64349-P). J.A. is supported through grant no. SEV-2013-0347. P.C. is supported through the European Union's YEI funds.

## Endnote

<sup>1</sup>In [23,101] a slightly different form of this equation was used, with the matrix product ( $\mathbf{F} \mathbf{G}$ ) instead of ( $\mathbf{G} \mathbf{F}$ ) in the second term on the right. While the former can be interpreted as a filtering criteria for the stability of mutants (fitness is applied once mutation has occurred), the latter represents more precisely the evolutionary process here described (fitness affects replication rates and mutation occurs concomitantly). The qualitative results of both expressions are equivalent, in practice only differing in the specific moment when the population state is measured.

## References

1. Lyell C. 1830 *Principles of geology, being an attempt to explain the former changes of the Earth's surface, by reference to causes now in operation*. London, UK: John Murray.
2. Darwin C. 1859 *On the origin of species by means of natural selection, or the preservation of favoured races in the struggle for life*, 1st edn. London, UK: John Murray.
3. Eldredge N, Gould SJ. 1972 Punctuated equilibria an alternative to phyletic gradualism. In *Models in paleobiology* (ed. TJM Schopf), pp. 82–115. San Francisco: Freeman Cooper.
4. Solé RV, Manrubia SC. 1996 Extinction and self-organized criticality in a model of large-scale evolution. *Phys. Rev. E* **54**, R42. (doi:10.1103/PhysRevE.54.R42)
5. Solé RV, Manrubia SC, Benton MJ, Bak P. 1997 Self-similarity of extinction statistics in the fossil record. *Nature* **388**, 764–767. (doi:10.1038/41996)
6. Hunt G, Hopkins MJ, Lidgard S. 2012 Simple versus complex models of trait evolution and stasis as a response to environmental change. *Proc. Natl Acad. Sci. USA* **112**, 4885–4890. (doi:10.1073/pnas.1403662111)
7. Solé RV, Manrubia SC, Benton MJ, Kauffman S, Bak P. 1999 Criticality and scaling in evolutionary ecology. *Trends Ecol. Evol.* **14**, 156–160. (doi:10.1016/S0169-5347(98)01518-3)
8. Scheffer M, Carpenter S, Foley JA, Folke C, Walker B. 2001 Catastrophic shifts in ecosystems. *Nature* **413**, 591–596. (doi:10.1038/35098000)
9. Foley JA, Coe MT, Scheffer M, Wang G. 2003 Regime shifts in the Sahara and Sahel: interactions between ecological and climatic systems in Northern Africa. *Ecosystems* **6**, 524–532. (doi:10.1007/s10021-002-0227-0)
10. May RM. 1977 Thresholds and breakpoints in ecosystems with a multiplicity of stable states. *Nature* **269**, 471–477. (doi:10.1038/269471a0)
11. Barnosky AD *et al.* 2012 Approaching a state shift in Earth's biosphere. *Nature* **486**, 52–58. (doi:10.1038/nature11018)
12. Benton MJ. 1993 *The fossil record 2*. London, UK: Chapman & Hall.
13. Harland WB, Armstrong RL, Cox AV, Craig LE, Smith AG, Smith DG. 1990 *A Geologic time scale 1989*. Cambridge, UK: Cambridge University Press.
14. Scheffer M *et al.* 2009 Early-warning signals for critical transitions. *Nature* **461**, 53–59. (doi:10.1038/nature08227)
15. Dai L, Vorselen D, Korolev KS, Gore J. 2012 Generic indicators for loss of resilience before a tipping point leading to population collapse. *Science* **336**, 1175–1177. (doi:10.1126/science.1219805)
16. Meijer M. 2000 *Bio-manipulation in the Netherlands: 15 years of experience*. Wageningen, The Netherlands: Wageningen University.
17. Kassas M. 1995 Desertification: a general review. *J. Arid Environ.* **30**, 115–128. (doi:10.1016/S0140-1963(05)80063-1)
18. Scheffer M, Hosper S, Meijer M-L, Moss B, Jeppesen E. 1993 Alternative equilibria in shallow lakes. *Trends Ecol. Evol.* **8**, 275–279. (doi:10.1016/0169-5347(93)90254-M)
19. Dublin HT, Sinclair A, McGlade J. 1990 Elephants and fire as causes of multiple stable states in the Serengeti-Mara woodlands. *J. Anim. Ecol.* **59**, 1147–1164. (doi:10.2307/5037)
20. Levin SA. 1992 The problem of pattern and scale in ecology: the Robert H. MacArthur Award lecture. *Ecology* **73**, 1943–1967. (doi:10.2307/1941447)
21. Stadler BMR, Stadler PF, Wagner GP, Fontana W. 2001 The topology of the possible: formal spaces underlying patterns of evolutionary change. *J. Theor. Biol.* **213**, 241–274. (doi:10.1006/jtbi.2001.2423)
22. Huynen MA, Stadler PF, Fontana W. 1996 Smoothness within ruggedness: the role of neutrality in adaptation. *Proc. Natl Acad. Sci. USA* **93**, 397–401. (doi:10.1073/pnas.93.1.397)
23. Aguirre J, Manrubia S. 2015 Tipping points and early warning signals in the genomic composition of populations induced by environmental changes. *Sci. Rep.* **5**, 9664. (doi:10.1038/srep09664)
24. Koelle K, Cobey S, Grenfell B, Pascual M. 2006 Epochal evolution shapes the phylodynamics of inter-pandemic influenza A (H3N2) in humans. *Science* **314**, 1898–1903. (doi:10.1126/science.1132745)
25. Wolf YI, Viboud C, Holmes EC, Koonin EV, Lipman DJ. 2006 Long intervals of stasis punctuated by bursts of positive selection in the seasonal evolution of influenza A virus. *Biol. Direct* **1**, 34. (doi:10.1186/1745-6150-1-34)
26. Wright S. 1931 Evolution in Mendelian populations. *Genetics* **16**, 97–159.
27. Svensson EI, Calsbeek R. 2012 *The adaptive landscape in evolutionary biology*. Oxford, UK: Oxford University Press.
28. Wagner A. 2011 *The origins of evolutionary innovations*. Oxford, UK: Oxford University Press.
29. Conant GC, Wolfe KH. 2008 Turning a hobby into a job: how duplicated genes find new functions. *Nat. Rev. Genet.* **9**, 938–950. (doi:10.1038/nrg2482)

30. Catalán P, Arias CF, Cuesta JA, Manrubia S. 2017 Adaptive multiscapes: an up-to-date metaphor to visualize molecular adaptation. *Biol. Direct* **12**, 7. (doi:10.1186/s13062-017-0178-1)
31. Hinkley T, Martins J, Chappay C, Haddad M, Stawiski E, Whitcomb JM, Petropoulos CJ, Bonhoeffer S. 2011 A systems analysis of mutational effects in HIV-1 protease and reverse transcriptase. *Nat. Genet.* **43**, 487–489. (doi:10.1038/ng.795)
32. Payne JL, Wagner A. 2014 The robustness and evolvability of transcription factor binding sites. *Science* **343**, 875–877. (doi:10.1126/science.1249046)
33. Steinberg B, Ostermeier M. 2016 Environmental changes bridge evolutionary valleys. *Sci. Adv.* **2**, e1500921. (doi:10.1126/sciadv.1500921)
34. Waddington CH. 1953 Genetic assimilation of an acquired character. *Evolution* **7**, 118–126. (doi:10.1111/j.1558-5646.1953.tb00070.x)
35. Kimura M. 1968 Evolutionary rate at the molecular level. *Nature* **217**, 624–626. (doi:10.1038/217624a0)
36. Kimura M. 1984 *The neutral theory of molecular evolution*. Cambridge, UK: Cambridge University Press.
37. Maynard Smith J. 1970 Natural selection and the concept of a protein space. *Nature* **225**, 563–564. (doi:10.1038/225563a0)
38. Schuster P, Fontana W, Stadler PF, Hofacker IL. 1994 From sequences to shapes and back: a case study in RNA secondary structures. *Proc. R. Soc. Lond. B* **255**, 279–284. (doi:10.1098/rspb.1994.0040)
39. Bastolla U, Porto M, Roman HE, Vendruscolo M. 2003 Connectivity of neutral networks, overdispersion, and structural conservation in protein evolution. *J. Mol. Biol.* **56**, 243–254. (doi:10.1007/s00239-002-2350-0)
40. Ciliberti S, Martin OC, Wagner A. 2007 Innovation and robustness in complex regulatory gene networks. *Proc. Natl Acad. Sci. USA* **104**, 13 595–13 596. (doi:10.1073/pnas.0705396104)
41. Matias Rodrigues JF, Wagner A. 2011 Genotype networks, innovation, and robustness in sulfur metabolism. *BMC Syst. Biol.* **5**, 39. (doi:10.1186/1752-0509-5-39)
42. Schultes EA, Bartel DP. 2000 One sequence, two ribozymes: implications for the emergence of new ribozyme folds. *Science* **289**, 448–452. (doi:10.1126/science.289.5478.448)
43. Bloom JD, Romero PA, Lu Z, Arnold FH. 2007 Neutral genetic drift can alter promiscuous protein functions, potentially aiding functional evolution. *Biol. Dir.* **2**, 17. (doi:10.1186/1745-6150-2-17)
44. Aguilar-Rodríguez J, Payne JL, Wagner A. 2017 A thousand empirical adaptive landscapes and their navigability. *Nat. Ecol. Evol.* **1**, 45. (doi:10.1038/s41559-016-0045)
45. Ohta T. 1973 Slightly deleterious mutant substitutions in evolution. *Nature* **246**, 96–98. (doi:10.1038/246096a0)
46. Eigen M. 1971 Selforganization of matter and the evolution of biological macromolecules. *Naturwissenschaften* **58**, 465–523. (doi:10.1007/BF00623322)
47. Domingo E (ed.). 2006 *Quasispecies: concept and implications for virology*. Berlin, Germany: Springer.
48. Woo H-J, Reifman J. 2012 A quantitative quasispecies theory-based model of virus escape mutation under immune selection. *Proc. Natl Acad. Sci. USA* **109**, 12 980–12 985. (doi:10.1073/pnas.1117201109)
49. Ferrada E, Wagner A. 2012 A comparison of genotype–phenotype maps for RNA and proteins. *Biophys. J.* **102**, 1916–1925. (doi:10.1016/j.bpj.2012.01.047)
50. Dingle K, Schaper S, Louis AA. 2015 The structure of the genotype–phenotype map strongly constrains the evolution of non-coding RNA. *J. R. Soc. Interface* **5**, 20150053. (doi:10.1098/rsfs.2015.0053)
51. Irbäck A, Troein C. 2002 Enumerating designing sequences in the HP model. *J. Biol. Phys.* **28**, 1–15. (doi:10.1023/A:1016225010659)
52. Garcia-Martin JA, Catalán P, Manrubia S, Cuesta JA. Statistical theory of phenotype abundance distributions: a test through exact enumeration of genotype spaces. <https://arxiv.org/abs/1806.03885>
53. Arias CF, Catalán P, Manrubia S, Cuesta JA. 2014 toyLIFE: a computational framework to study the multi-level organisation of the genotype–phenotype map. *Sci. Rep.* **4**, 7549. (doi:10.1038/srep07549)
54. Greenbury S, Ahnert S. 2015 The organization of biological sequences into constrained and unconstrained parts determines fundamental properties of genotype–phenotype maps. *J. R. Soc. Interface* **12**, 20150724. (doi:10.1098/rsif.2015.0724)
55. Manrubia S, Cuesta JA. 2017 Distribution of genotype network sizes in sequence-to-structure genotype–phenotype maps. *J. R. Soc. Interface* **14**, 20160976. (doi:10.1098/rsif.2016.0976)
56. Hofacker IL, Fontana W, Stadler PF, Bonhoeffer LS, Tacker M, Schuster P. 1994 Fast folding and comparison of RNA secondary structures. *Monatshfte f. Chemie* **125**, 167–188. (doi:10.1007/BF00818163)
57. Grüner W, Giegerich R, Strothmann D, Reidys C, Weber J, Hofacker IL, Stadler PF, Schuster P. 1996 Analysis of RNA sequence structure maps by exhaustive enumeration. I. Neutral networks. *Monatshfte f. Chemie* **127**, 355–374. (doi:10.1007/BF00810881)
58. Fontana W, Schuster P. 1998 Continuity in evolution: on the nature of transitions. *Science* **280**, 1451–1455. (doi:10.1126/science.280.5368.1451)
59. Cowperthwaite MC, Economo EP, Harcombe WR, Miller EL, Meyers LA. 2008 The ascent of the abundant: how mutational networks constrain evolution. *PLoS Comput. Biol.* **4**, e1000110. (doi:10.1371/journal.pcbi.1000110)
60. Jörg T, Martin OC, Wagner A. 2008 Neutral network sizes of biological RNA molecules can be computed and are not atypically small. *BMC Bioinformatics* **9**, 464. (doi:10.1186/1471-2105-9-464)
61. Aguirre J, Buldú JM, Stich M, Manrubia SC. 2011 Topological structure of the space of phenotypes: the case of RNA neutral networks. *PLoS ONE* **6**, e26324. (doi:10.1371/journal.pone.0026324)
62. Lau KF, Dill KA. 1989 A lattice statistical mechanics model of the conformational and sequence spaces of proteins. *Macromolecules* **22**, 3986–3997. (doi:10.1021/ma00200a030)
63. Lipman DJ, Wilbur WJ. 1991 Modelling neutral and selective evolution of protein folding. *Proc. R. Soc. Lond. B* **245**, 7–11. (doi:10.1098/rspb.1991.0081)
64. Li H, Helling R, Tang C, Wingreen N. 1996 Emergence of preferred structures in a simple model of protein folding. *Science* **273**, 666–669. (doi:10.1126/science.273.5275.666)
65. Bornberg-Bauer E. 1997 How are model protein structures distributed in sequence space? *Biophys. J.* **73**, 2393–2403. (doi:10.1016/S0006-3495(97)78268-7)
66. Crombach A. 2017 *Modelling the evolution of dynamic regulatory networks: some critical insights*, pp. 257–273. Cham, Switzerland: Springer International Publishing.
67. Payne JL, Moore JH, Wagner A. 2014 Robustness, evolvability, and the logic of genetic regulation. *Artif. Life* **20**, 111–126. (doi:10.1162/ARTL\_a\_00099)
68. Matias Rodrigues JF, Wagner A. 2009 Evolutionary plasticity and innovations in complex metabolic reaction networks. *PLoS Comp. Biol.* **5**, e1000613. (doi:10.1371/journal.pcbi.1000613)
69. Barve A, Wagner A. 2013 A latent capacity for evolutionary innovation through exaptation in metabolic systems. *Nature* **500**, 203–206. (doi:10.1038/nature12301)
70. Hosseini S-R, Barve A, Wagner A. 2015 Exhaustive analysis of a genotype space comprising 10<sup>15</sup> central carbon metabolisms reveals an organization conducive to metabolic innovation. *PLoS Comput. Biol.* **11**, e1004329. (doi:10.1371/journal.pcbi.1004329)
71. Ibáñez-Marcelo E, Alarcón T. 2014 The topology of robustness and evolvability in evolutionary systems with genotype–phenotype map. *J. Theor. Biol.* **356**, 144–162. (doi:10.1016/j.jtbi.2014.04.014)
72. Johnston IG, Ahnert SE, Doye JP, Louis AA. 2011 Evolutionary dynamics in a simple model of self-assembly. *Phys. Rev. E* **83**, 066105. (doi:10.1103/PhysRevE.83.066105)
73. Greenbury SF, Johnston IG, Louis AA, Ahnert SE. 2014 A tractable genotype–phenotype map for the self-assembly of protein quaternary structure. *J. R. Soc. Interface* **11**, 20140249. (doi:10.1098/rsif.2014.0249)
74. Catalán P, Wagner A, Manrubia S, Cuesta JA. 2018 Adding levels of complexity enhances robustness and evolvability in a multi-level genotype–phenotype map. *J. R. Soc. Interface* **15**, 20170516. (doi:10.1098/rsif.2017.0516)
75. Ahnert SE. 2017 Structural properties of genotype–phenotype maps. *J. R. Soc. Interface* **14**, 20170275. (doi:10.1098/rsif.2017.0275)
76. Khatri BS, McLeish TCB, Sear RP. 2009 Statistical mechanics of convergent evolution in spatial patterning. *Proc. Natl Acad. Sci. USA* **106**, 9564–9569. (doi:10.1073/pnas.0812260106)

77. Fontana W, Schuster P. 1998 Shaping space: the possible and the attainable in RNA genotype–phenotype mapping. *J. Theor. Biol.* **194**, 491–515. (doi:10.1006/jtbi.1998.0771)
78. Schaper S, Louis AA. 2014 The arrival of the frequent: how bias in genotype–phenotype maps can steer populations to local optima. *PLoS ONE* **9**, e86635. (doi:10.1371/journal.pone.0086635)
79. Manrubia S, Cuesta JA. 2015 Evolution on genotype networks accelerates the ticking rate of the molecular clock. *J. R. Soc. Interface* **12**, 20141010. (doi:10.1098/rsif.2014.1010)
80. Greenbury SF, Schaper S, Ahnert SE, Louis AA. 2016 Genetic correlations greatly increase mutational robustness and can both reduce and enhance evolvability. *PLoS Comput. Biol.* **12**, e1004773. (doi:10.1371/journal.pcbi.1004773)
81. Bornberg-Bauer E, Chan HS. 1999 Modeling evolutionary landscapes: mutational stability, topology, and superfunnels in sequence space. *Proc. Natl Acad. Sci. USA* **96**, 10 689–10 694. (doi:10.1073/pnas.96.19.10689)
82. Wuchty S, Fontana W, Hofacker IL, Schuster P. 1999 Complete suboptimal folding of RNA and the stability of secondary structures. *Biopolymers* **49**, 145–165. (doi:10.1002/(SICI)1097-0282(199902)49:2<145::AID-BIP4>3.0.CO;2-G)
83. Ancel LW, Fontana W. 2000 Plasticity, evolvability, and modularity in RNA. *J. Exp. Zool.* **288**, 242–283. (doi:10.1002/1097-010X(20001015)288:3<242::AID-JEZ5>3.0.CO;2-O)
84. Grüner W, Giegerich R, Strothmann D, Reidys C, Weber J, Hofacker IL, Stadler PF, Schuster P. 1996 Analysis of RNA sequence structure maps by exhaustive enumeration II. Structures of neutral networks and shape space covering. *Monatshefte f. Chemie* **127**, 375–389. (doi:10.1007/BF00810882)
85. Kivelä M, Arenas A, Barthélemy M, Gleeson JP, Moreno Y, Porter MA. 2010 Multilayer networks. *J. Comp. Net.* **2**, 203–271. (doi:10.1093/comnet/cnu016)
86. Boccaletti S, Bianconi G, Criado R, del Genio C, Gómez-Gardeñes J, Romance M, Sendiña-Nadal I, Wang Z, Zanin M. 2014 The structure and dynamics of multilayer networks. *Phys. Rep.* **544**, 1–122. (doi:10.1016/j.physrep.2014.07.001)
87. Manrubia SC. 2012 Modelling viral evolution and adaptation: challenges and rewards. *Curr. Opin. Virol.* **2**, 531–537. (doi:10.1016/j.coviro.2012.06.006)
88. Li H, Tang C, Wingreen NS. 2002 Designability of protein structures: A lattice-model study using the Miyazawa-Jernigan matrix. *Proteins* **49**, 403–412. (doi:10.1002/prot.10239)
89. Stich M, Lázaro E, Manrubia SC. 2010 Phenotypic effect of mutations in evolving populations of RNA molecules. *BMC Evol. Biol.* **10**, 46. (doi:10.1186/1471-2148-10-46)
90. Poelwijk FJ, Kiviet DJ, Weinreich DM, Tans SJ. 2007 Empirical fitness landscapes reveal accessible evolutionary paths. *Nature* **445**, 383–386. (doi:10.1038/nature05451)
91. Jiménez JI, Xulvi-Brunet R, Campbell GW, Turk-MacLeod R, Chen IA. 2013 Comprehensive experimental fitness landscape and evolutionary network for small RNA. *Proc. Natl Acad. Sci. USA* **110**, 14 984–14 989. (doi:10.1073/pnas.1307604110)
92. De Visser JAG, Krug J. 2014 Empirical fitness landscapes and the predictability of evolution. *Nat. Rev. Genet.* **15**, 480. (doi:10.1038/nrg3744)
93. De Vos MG, Dawid A, Sunderlikova V, Tans SJ. 2015 Breaking evolutionary constraint with a tradeoff ratchet. *Proc. Natl Acad. Sci. USA* **112**, 14 906–14 911. (doi:10.1073/pnas.1510282112)
94. Firmberg E, Labonte JW, Gray JJ, Ostermeier M. 2014 A comprehensive, high-resolution map of a genes fitness landscape. *Mol. Biol. Evol.* **31**, 1581–1592. (doi:10.1093/molbev/msu081)
95. Lauring AS, Andino R. 2011 Exploring the fitness landscape of an RNA virus by using a universal barcode microarray. *J. Virol.* **85**, 3780–3791. (doi:10.1128/JVI.02217-10)
96. Kouyos RD, Leventhal GE, Hinkley T, Haddad M, Whitcomb JM, Petropoulos CJ, Bonhoeffer S. 2012 Exploring the complexity of the HIV-1 fitness landscape. *PLoS Genet.* **8**, e1002551. (doi:10.1371/journal.pgen.1002551)
97. Bank C, Matuszewski S, Hietpas RT, Jensen JD. 2016 On the (un)predictability of a large intragenic fitness landscape. *Proc. Natl Acad. Sci. USA* **113**, 14 085–14 090. (doi:10.1073/pnas.1612676113)
98. Zagorski M, Burda Z, Waclaw B. 2016 Beyond the hypercube: evolutionary accessibility of fitness landscapes with realistic mutational networks. *PLoS Comp. Biol.* **12**, e1005218. (doi:10.1371/journal.pcbi.1005218)
99. Kauffman S, Levin S. 1987 Towards a general theory of adaptive walks on rugged landscapes. *J. Theor. Biol.* **128**, 11–45. (doi:10.1016/S0022-5193(87)80029-2)
100. Østman B, Adami C. 2014 Predicting evolution and visualizing high-dimensional fitness landscapes. In *Recent advances in the theory and application of fitness landscapes* (eds H Richter, A Engelbrecht), pp. 509–526. Berlin, Germany: Springer.
101. Yubero P, Manrubia S, Aguirre J. 2017 The space of genotypes is a network of networks: implications for evolutionary and extinction dynamics. *Sci. Rep.* **7**, 13813. (doi:10.1038/s41598-017-14048-x)
102. Wilke CO. 2001 Adaptive evolution on neutral networks. *Bull. Math. Biol.* **63**, 715–730. (doi:10.1006/bulm.2001.0244)
103. Aguirre J, Buldú JM, Manrubia SC. 2009 Evolutionary dynamics on networks of selectively neutral genotypes: effects of topology and sequence stability. *Phys. Rev. E* **80**, 066112. (doi:10.1103/PhysRevE.80.066112)
104. Boccaletti S, Latora V, Moreno Y, Chavez M, Hwang DU. 2006 Complex networks: structure and dynamics. *Phys. Rep.* **424**, 175–308. (doi:10.1016/j.physrep.2005.10.009)
105. Reeves T, Farr RS, Blundell J, Gallagher A, Fink TMA. 2016 Eigenvalues of neutral networks: interpolating between hypercubes. *Discrete. Math.* **339**, 1283–1290. (doi:10.1016/j.disc.2015.11.011)
106. van Nimwegen E, Crutchfield JP, Huynen M. 1999 Neutral evolution of mutational robustness. *Proc. Natl Acad. Sci. USA* **96**, 9716–9720. (doi:10.1073/pnas.96.17.9716)
107. Wagner A. 2008 Robustness and evolvability: a paradox resolved. *Proc. R. Soc. B* **275**, 91–100. (doi:10.1098/rspb.2007.1137)
108. Draghi JA, Parsons TL, Wagner GP, Plotkin JB. 2010 Mutational robustness can facilitate adaptation. *Nature* **463**, 353–355. (doi:10.1038/nature08694)
109. Reidys C, Stadler PF, Schuster P. 1997 Generic properties of combinatorial maps: neutral networks of RNA secondary structures. *Bull. Math. Biol.* **59**, 339–397. (doi:10.1007/BF02462007)
110. Fontana W. 2002 Modelling ‘evo-devo’ with RNA. *BioEssays* **24**, 1164–1177. (doi:10.1002/bies.10190)
111. Zuckerkandl E, Pauling L. 1965 Evolutionary divergence and convergence in proteins. In *Evolving genes and proteins* (eds V Bryson, H Vogel), pp. 97–166. New York, NY: Academic Press.
112. Takahata N. 1987 On the overdispersed molecular clock. *Genetics* **116**, 169–179.
113. Aguirre J, Papo D, Buldú JM. 2013 Successful strategies for competing networks. *Nat. Phys.* **9**, 230–234. (doi:10.1038/nphys2556)
114. Buldú JM, Sevilla-Escoboza R, Aguirre J, Papo D, Gutiérrez R. 2016 Interconnecting networks: the role of connector links. In *Interconnected networks* (ed. A Garas), pp. 61–77. Berlin, Germany: Springer International Publishing.
115. Irazo J, Buldú JM, Aguirre J. 2016 Competition among networks highlights the power of the weak. *Nat. Comm.* **7**, 13273. (doi:10.1038/ncomms13273)
116. Newman MEJ. 2010 *Networks: an introduction*. New York, NY: Oxford University Press.
117. Langville C, Meyer AN. 2006 *Google’s pagerank and beyond: the science of search engine rankings*. Princeton, NJ: Princeton University Press.
118. Senanayake U, Piraveenan M, Zomaya A. 2015 The pagerank-index: going beyond citation counts in quantifying scientific impact of researchers. *PLoS ONE* **10**, e0134794. (doi:10.1371/journal.pone.0134794)
119. Bergstrom C. 2007 Eigenfactor: measuring the value and prestige of scholarly journals. *C&RL News* **68**, 314–316. (doi:10.5860/crln.68.5.7804)
120. Seary AJ, Richards WD. 2003 Spectral methods for analyzing and visualizing networks: an introduction. In *Dynamic social network modeling and analysis: workshop summary and papers*, pp. 209–228. Washington, DC: The National Academies Press.
121. Lohmann G *et al.* 2010 Eigenvector centrality mapping for analyzing connectivity patterns in fMRI data of the human brain. *PLoS ONE* **5**, e10232. (doi:10.1371/journal.pone.0010232)
122. Capitán JA, Aguirre J, Manrubia S. 2015 Dynamical community structure of populations evolving on genotype networks. *Chaos, Solitons Fractals* **72**, 99–106. (doi:10.1016/j.chaos.2014.11.019)
123. van Nimwegen E, Crutchfield JP. 2000 Metastable evolutionary dynamics: crossing fitness barriers or escaping via neutral paths? *Bull. Math. Biol.* **62**, 799–848. (doi:10.1006/bulm.2000.0180)
124. Wilke CO, Wang JL, Ofria C, Lenski RE, Adami C. 2001 Evolution of digital organisms at high

- mutation rates leads to survival of the flattest. *Nature* **412**, 331–333. (doi:10.1038/35085569)
125. Codo ner FM, Darós J-A, Solé RV, Elena SF. 2006 The fittest versus the flattest: experimental confirmation of the quasispecies effect with subviral pathogens. *PLoS Pathog.* **2**, e136. (doi:10.1371/journal.ppat.0020136)
126. Wilke CO, Adami C. 2003 Evolution of mutational robustness. *Mut. Res.* **522**, 3–11. (doi:10.1016/S0027-5107(02)00307-X)
127. Wilke CO. 2001 Selection for fitness versus selection for robustness in RNA secondary structure folding. *Evolution* **55**, 2412–2420. (doi:10.1111/j.0014-3820.2001.tb00756.x)
128. 2008 Simple quasispecies models for the survival-of-the-flattest effect: the role of space. *J. Theor. Biol.* **250**, 560–568. (doi:10.1016/j.jtbi.2007.10.027)
129. Colizzi ES, Hogeweg P. 2014 Evolution of functional diversification within quasispecies. *Genome Biol. Evol.* **6**, 1990–2007. (doi:10.1093/gbe/evu150)
130. Beardmore RE, Gudelj I, Lipson DA, Hurst LD. 2011 Metabolic trade-offs and the maintenance of the fittest and the flattest. *Nature* **472**, 342–346. (doi:10.1038/nature09905)
131. van Nimwegen E, Crutchfield JP. 2000 Optimizing epochal evolutionary search: population-size independent theory. *Comput. Methods Appl. Mech. Eng.* **186**, 171–194. (doi:10.1016/S0045-7825(99)00383-7)
132. van Nimwegen E, Crutchfield JP. 2001 Optimizing epochal evolutionary search: population-size dependent theory. *Mach. Learn.* **45**, 77–114. (doi:10.1023/A:1010928206141)
133. Fuentes-Hernandez A, Pluacain J, Gori F, Pena-Miller R, Reding C, Jansen G, Schulenburg H, Gudelj I, Beardmore R. 2015 Using a sequential regimen to eliminate bacteria at sublethal antibiotic dosages. *PLoS Biol.* **13**, e1002104. (doi:10.1371/journal.pbio.1002104)
134. Gavrilets S. 2004 *Fitness landscapes and the origin of species*. Princeton, NJ: Princeton University Press.
135. Buldyrev SV, Parshani R, Gerald P, Stanley HE, Havlin S. 2010 Catastrophic cascade of failures in interdependent networks. *Nature* **464**, 1025–1028. (doi:10.1038/nature08932)
136. Gao J, Buldyrev SV, Havlin S, Stanley HE. 2011 Robustness of a network of networks. *Phys. Rev. Lett.* **107**, 195701. (doi:10.1103/PhysRevLett.107.195701)
137. Um J, Minnhagen P, Kim BJ. 2011 Synchronization in interdependent networks. *Chaos* **21**, 025106. (doi:10.1063/1.3596698)
138. Aguirre J, Sevilla-Escoboza R, Gutiérrez R, Papo D, Buldú JM. 2014 Synchronization of interconnected networks: the role of connector nodes. *Phys. Rev. Lett.* **112**, 248701. (doi:10.1103/PhysRevLett.112.248701)
139. Gómez-Gardeñes J, Reinares I, Arenas A, Floria LM. 2012 Evolution of cooperation in multiplex networks. *Sci. Rep.* **2**, 620. (doi:10.1038/srep00620)
140. Wang Z, Szolnoki A, Perc M. 2012 Evolution of public cooperation on interdependent networks: the impact of biased utility functions. *Europhys. Lett.* **97**, 48001. (doi:10.1209/0295-5075/97/48001)
141. Dickison M, Havlin S, Stanley HE. 2012 Epidemics on interconnected networks. *Phys. Rev. E* **85**, 066109. (doi:10.1103/PhysRevE.85.066109)
142. Saumell-Mendiola A, Serrano MA, Boguñá M. 2012 Epidemic spreading on interconnected networks. *Phys. Rev. E* **86**, 026106. (doi:10.1103/PhysRevE.86.026106)
143. Yağan O, Gligor V. 2012 Analysis of complex contagions in random multiplex networks. *Phys. Rev. E* **86**, 036103. (doi:10.1103/PhysRevE.86.036103)
144. O'Brien PJ, Herschlag D. 1999 Catalytic promiscuity and the evolution of new enzymatic activities. *Chem. Biol.* **6**, R91–R105. (doi:10.1016/S1074-5521(99)80033-7)
145. Copley SD. 2003 Enzymes with extra talents: moonlighting functions and catalytic promiscuity. *Curr. Opin. Chem. Biol.* **7**, 265–272. (doi:10.1016/S1367-5931(03)00032-2)
146. Babtie A, Tokuriki N, Hoffelder F. 2010 What makes an enzyme promiscuous? *Curr. Opin. Chem. Biol.* **14**, 200–207. (doi:10.1016/j.cbpa.2009.11.028)
147. Khersonsky O, Tawfik DS. 2010 Enzyme promiscuity: a mechanistic and evolutionary perspective. *Annu. Rev. Biochem.* **79**, 471–505. (doi:10.1146/annurev-biochem-030409-143718)
148. Copley SD. 2015 An evolutionary biochemist's perspective on promiscuity. *Trends Biochem. Sci.* **40**, 72–78. (doi:10.1016/j.tibs.2014.12.004)
149. Copley SD. 2017 Shining a light on enzyme promiscuity. *Curr. Opin. Struct. Biol.* **47**, 167–175. (doi:10.1016/j.sbi.2017.11.001)
150. Schmidt DMZ, Mundorff EC, Dojka M, Bermudez E, Ness JE, Govindarajan S, Babbitt PC, Minshall J, Gerlt JA. 2003 Evolutionary potential of ( $\beta/\alpha$ ) 8-barrels: functional promiscuity produced by single substitutions in the enolase superfamily. *Biochemistry* **42**, 8387–8393. (doi:10.1021/bi034769a)
151. Aharoni A, Gaidukov L, Khersonsky O, Gould SM, Roodveldt C, Tawfik DS. 2005 The 'evolability' of promiscuous protein functions. *Nat. Genet.* **37**, 73. (doi:10.1038/ng1482)
152. Amitai G, Gupta RD, Tawfik DS. 2007 Latent evolutionary potentials under the neutral mutational drift of an enzyme. *HFSP J.* **1**, 67–78. (doi:10.2976/1.2739115/10.2976/1)
153. Bloom JD, Raval A, Wilke CO. 2007 Thermodynamics of neutral protein evolution. *Genetics* **175**, 255–266. (doi:10.1534/genetics.106.061754)
154. Yang G, Hong N, Baier F, Jackson CJ, Tokuriki N. 2016 Conformational tinkering drives evolution of a promiscuous activity through indirect mutational effects. *Biochemistry* **55**, 4583–4593. (doi:10.1021/acs.biochem.6b00561)
155. Baier F, Hong N, Yang G, Pabis A, Barrozo A, Carr PD, Kamerlin SC, Jackson CJ, Tokuriki N. 2017 Cryptic genetic variation defines the adaptive evolutionary potential of enzymes. *bioRxiv*. (doi:10.1101/232793)
156. Paaby AB, Rockman MV. 2014 Cryptic genetic variation: evolution's hidden substrate. *Nat. Rev. Gen.* **15**, 247–258. (doi:10.1038/nrg3688)
157. Hughes AL. 1994 The evolution of functionally novel proteins after gene duplication. *Proc. R. Soc. Lond. B* **256**, 119–124. (doi:10.1098/rspb.1994.0058)
158. Kim J, Kershner JP, Novikov Y, Shoemaker RK, Copley SD. 2010 Three serendipitous pathways in *E. coli* can bypass a block in pyridoxal-5'-phosphate synthesis. *Mol. Syst. Biol.* **6**, 436. (doi:10.1038/msb.2010.88)
159. Bratulic S, Gerber F, Wagner A. 2015 Mistranslation drives the evolution of robustness in tem-1  $\beta$ -lactamase. *Proc. Natl Acad. Sci. USA* **112**, 12758–12763. (doi:10.1073/pnas.1510071112)
160. Whitehead DJ, Wilke CO, Vernazobres D, Bornberg-Bauer E. 2008 The look-ahead effect of phenotypic mutations. *Biol. Direct* **3**, 18. (doi:10.1186/1745-6150-3-18)
161. Drummond DA, Wilke CO. 2009 The evolutionary consequences of erroneous protein synthesis. *Nat. Rev. Genet.* **10**, 715–724. (doi:10.1038/nrg2662)
162. Yanagida H, Gispán A, Kadouri N, Rozen S, Sharon M, Barkai N, Tawfik DS. 2015 The evolutionary potential of phenotypic mutations. *PLoS Genet.* **11**, e1005445. (doi:10.1371/journal.pgen.1005445)
163. Vaidya N, Lehman N. 2009 One RNA plays three roles to provide catalytic activity to a group I intron lacking an endogenous internal guide sequence. *Nucleic Acids Res.* **37**, 3981–3989. (doi:10.1093/nar/gkp271)
164. Talini G, Branciamore S, Gallori E. 2011 Ribozymes: flexible molecular devices at work. *Biochimie* **93**, 1998–2005. (doi:10.1016/j.biochi.2011.06.026)
165. Lau MW, Unrau PJ. 2009 A promiscuous ribozyme promotes nucleotide synthesis in addition to ribose chemistry. *Chem. Biol.* **16**, 815–825. (doi:10.1016/j.chembiol.2009.07.005)
166. Hayden EJ, Ferrada E, Wagner A. 2011 Cryptic genetic variation promotes rapid evolutionary adaptation in an RNA enzyme. *Nature* **474**, 92–95. (doi:10.1038/nature10083)
167. Wagner A. 2014 Mutational robustness accelerates the origin of novel RNA phenotypes through phenotypic plasticity. *Biophys. J.* **106**, 955–965. (doi:10.1016/j.bpj.2014.01.003)
168. Elowitz MB, Levine AJ, Siggia ED, Swain PS. 2002 Stochastic gene expression in a single cell. *Science* **297**, 1183–1186. (doi:10.1126/science.1070919)
169. Raser JM, O'Shea EK. 2005 Noise in gene expression: origins, consequences, and control. *Science* **309**, 2010–2013. (doi:10.1126/science.1105891)
170. Maheshri N, O'Shea EK. 2007 Living with noisy genes: how cells function reliably with inherent variability in gene expression. *Annu. Rev. Biophys. Biomol. Struct.* **36**, 413–434. (doi:10.1146/annurev.biophys.36.040306.132705)
171. Ackermann M. 2015 A functional perspective on phenotypic heterogeneity in microorganisms. *Nat. Rev. Microbiol.* **13**, 497–508. (doi:10.1038/nrmicro3491)

172. van Boxtel C, van Heerden JH, Nordholt N, Schmidt P, Bruggeman FJ. 2017 Taking chances and making mistakes: non-genetic phenotypic heterogeneity and its consequences for surviving in dynamic environments. *J. R. Soc. Interface* **14**, 20170141. (doi:10.1098/rsif.2017.0141)
173. Herranz H, Cohen SM. 2010 MicroRNAs and gene regulatory networks: managing the impact of noise in biological systems. *Gene Dev.* **24**, 1339–1344. (doi:10.1101/gad.1937010)
174. Lehner B. 2008 Selection to minimise noise in living systems and its implications for the evolution of gene expression. *Mol. Syst. Biol.* **4**, 170. (doi:10.1038/msb.2008.11)
175. Little SC, Tikhonov M, Gregor T. 2013 Precise developmental gene expression arises from globally stochastic transcriptional activity. *Cell* **154**, 789–800. (doi:10.1016/j.cell.2013.07.025)
176. Eldar A, Elowitz MB. 2010 Functional roles for noise in genetic circuits. *Nature* **467**, 167–173. (doi:10.1038/nature09326)
177. Veening J-W, Smits WK, Kuipers OP. 2008 Bistability, epigenetics, and bet-hedging in bacteria. *Annu. Rev. Microbiol.* **62**, 193–210. (doi:10.1146/annurev.micro.62.081307.163002)
178. Norman TM, Lord ND, Paulsson J, Losick R. 2015 Stochastic switching of cell fate in microbes. *Annu. Rev. Microbiol.* **69**, 381–403. (doi:10.1146/annurev-micro-091213-112852)
179. Süel GM, García-Ojalvo J, Liberman LM, Elowitz MB. 2006 An excitable gene regulatory circuit induces transient cellular differentiation. *Nature* **440**, 545–550. (doi:10.1038/nature04588)
180. Balaban NQ, Merrin J, Chait R, Kowalik L, Leibler S. 2004 Bacterial persistence as a phenotypic switch. *Science* **305**, 1622–1625. (doi:10.1126/science.1099390)
181. Beaumont HJ, Gallie J, Kost C, Ferguson GC, Rainey PB. 2009 Experimental evolution of bet hedging. *Nature* **462**, 90–93. (doi:10.1038/nature08504)
182. West-Eberhard MJ. 2003 *Developmental plasticity and evolution*. Oxford, UK: Oxford University Press.
183. Jeffery CJ. 1999 Moonlighting proteins. *Trends Biochem. Sci.* **24**, 8–11. (doi:10.1016/S0968-0004(98)01335-8)
184. Copley SD. 2012 Moonlighting is mainstream: paradigm adjustment required. *Bioessays* **34**, 578–588. (doi:10.1002/bies.201100191)
185. Piatigorsky J. 2007 *Gene sharing and evolution: the diversity of protein functions*. Cambridge, MA: Harvard University Press.
186. Kortmann J, Narberhaus F. 2012 Bacterial RNA thermometers: molecular zippers and switches. *Nat. Rev. Microbiol.* **10**, 255–265. (doi:10.1038/nrmicro2730)
187. García-Martin JA, Dotu I, Fernandez-Chamorro J, Lozano G, Ramajo J, Martínez-Salas E, Clote P. 2016 RNAiFold2T: constraint programming design of thermo-IRES switches. *Bioinformatics* **32**, i360–i368. (doi:10.1093/bioinformatics/btw265)
188. Payne JL, Wagner A. 2014 Latent phenotypes pervade gene regulatory circuits. *BMC Syst. Biol.* **8**, 64. (doi:10.1186/1752-0509-8-64)
189. Espinosa-Soto C, Martin OC, Wagner A. 2011 Phenotypic plasticity can facilitate adaptive evolution in gene regulatory circuits. *BMC Evol. Biol.* **11**, 5. (doi:10.1186/1471-2148-11-5)
190. Jiménez A, Cotterell J, Munteanu A, Sharpe J. 2017 A spectrum of modularity in multi-functional gene circuits. *Mol. Sys. Biol.* **13**, 925. (doi:10.15252/msb.20167347)
191. Via S, Lande R. 1985 Genotype–environment interaction and the evolution of phenotypic plasticity. *Evolution* **39**, 505–522. (doi:10.1111/j.1558-5646.1985.tb00391.x)
192. Scheiner SM. 1993 Genetics and evolution of phenotypic plasticity. *Annu. Rev. Ecol. Syst.* **24**, 35–68. (doi:10.1146/annurev.es.24.110193.000343)
193. Ancel LW. 2000 Undermining the Baldwin expediting effect: does phenotypic plasticity accelerate evolution? *Theor. Popul. Biol.* **58**, 307–319. (doi:10.1006/tpbi.2000.1484)
194. Price TD, Qvarnström A, Irwin DE. 2003 The role of phenotypic plasticity in driving genetic evolution. *Proc. R. Soc. Lond. B* **270**, 1433–1440. (doi:10.1098/rspb.2003.2372)
195. Lande R. 2009 Adaptation to an extraordinary environment by evolution of phenotypic plasticity and genetic assimilation. *J. Evol. Biol.* **22**, 1435–1446. (doi:10.1111/j.1420-9101.2009.01754.x)
196. Scheiner SM, Holt RD. 2012 The genetics of phenotypic plasticity. X. Variation versus uncertainty. *Ecol. Evol.* **2**, 751–767. (doi:10.1002/ece3.217)
197. Gómez-Mestre I, Jovani R. 2013 A heuristic model on the role of plasticity in adaptive evolution: plasticity increases adaptation, population viability and genetic variation. *Proc. R. Soc. B* **280**, 20131869. (doi:10.1098/rspb.2013.1869)
198. Draghi JA, Whitlock MC. 2012 Phenotypic plasticity facilitates mutational variance, genetic variance, and evolvability along the major axis of environmental variation. *Evolution* **66**, 2891–2902. (doi:10.1111/j.1558-5646.2012.01649.x)
199. McCaskill JS. 1990 The equilibrium partition function and base pair binding probabilities for RNA secondary structure. *Biopolymers* **29**, 1105–1119. (doi:10.1002/bip.360290621)
200. Koonin EV. 2011 Are there laws of genome evolution? *PLoS Comput. Biol.* **7**, e1002173. (doi:10.1371/journal.pcbi.1002173)
201. Irazo J, Gómez M, López de Saro F, Manrubia SC. 2014 Large-scale genomic analysis suggests a neutral punctuated dynamics of transposable elements in bacterial genomes. *PLoS Comput. Biol.* **10**, e1003680. (doi:10.1371/journal.pcbi.1003680)
202. Irazo J, Cuesta J, Manrubia S, Katsnelson M, Koonin E. 2017 Disentangling the effects of selection and loss bias on gene dynamics. *Proc. Natl Acad. Sci. USA* **114**, E5616. (doi:10.1073/pnas.1704925114)
203. Schönknecht G, Weber A, Lercher M. 2014 Horizontal gene acquisitions by eukaryotes as drivers of adaptive evolution. *BioEssays* **36**, 9–20. (doi:10.1002/bies.201300095)
204. Koonin EV. 2015 The turbulent network dynamics of microbial evolution and the statistical tree of life. *J. Mol. Evol.* **80**, 244–250. (doi:10.1007/s00239-015-9679-7)
205. Irazo J, Krupovic M, Koonin EV. 2016 The double-stranded DNA virosphere as a modular hierarchical network of gene sharing. *mBio* **7**, e00978. (doi:10.1128/mBio.00978-16)
206. Lucía-Sanz A, Manrubia S. 2017 Multipartite viruses: Adaptive trick or evolutionary treat? *npj Sys. Biol. App.* **3**, 34. (doi:10.1038/s41540-017-0035-y)
207. Ueda M, Takeuchi N, Kaneno K. 2017 Stronger selection can slow down evolution driven by recombination on a smooth fitness landscape. *PLoS ONE* **12**, e0183120. (doi:10.1371/journal.pone.0183120)
208. Peabody GL, Li H, Kao KC. 2017 Sexual recombination and increased mutation rate expedite evolution of *Escherichia coli* in varied fitness landscapes. *Nat. Commun.* **8**, 2112. (doi:10.1038/s41467-017-02323-4)
209. Cramer A, Raillard S, Bermudez E, Stemmer W. 1998 DNA shuffling of a family of genes from diverse species accelerates directed evolution. *Nature* **391**, 288–291. (doi:10.1038/34663)
210. Azevedo RB, Lohaus R, Srinivasan S, Dang KK, Burch CL. 2006 Sexual reproduction selects for robustness and negative epistasis in artificial gene networks. *Nature* **440**, 87–90. (doi:10.1038/nature04488)